# **Review Article** Vol. 21, No. 6, Jan.-Feb. 2026, p.

# Ascorbic Acid: Rationale and Applications in Inhibiting Enzymatic Browning of Fruits and Vegetables- A Comprehensive Review

O. Al-abbasy<sup>1\*</sup>, Z. Alsarraf<sup>2</sup>, N. Mahdi<sup>3</sup>, A. Al-barwari<sup>1</sup>

- 1- Department of Chemistry, College of Education for Pure Science, University of Mosul, Iraq
- (\*- Corresponding Author Email: chem.omar1978@uomosul.edu.iq)
- 2- Pharmaceutical Chemistry Department, College of Pharmacy, Ninevah University, Mosul, Iraq
- 3- General Directorate of Education in Nineveh, Iraq

Received: 11.12.2024 How to cite this article: Revised: 18.05.2025 Al-abbasy, O., Alsarraf, Z., Mahdi, N., & Al-barwari, A. (2025). Ascorbic Accepted: 18.06.2025 acid: Rationale and applications in inhibiting enzymatic browning of fruits Available Online: 02.12.2025 and vegetables- A comprehensive review. Iranian Food Science and Technology Research Journal. 21(6),. https://doi.org/10.22067/ifstrj.2025.91193.1391

### Abstract

Enzymatic browning mostly happens in fresh fruits and vegetables and is critical in determining the product's shelf life. A class of enzymes known as polyphenol oxidases are responsible for this color alteration. Polyphenol oxidase is the main enzyme that catalyzes the oxidation of phenolic compounds in the presence of oxygen, forming brown pigment. Therefore, several methods are required to prevent these reactions. Natural ascorbic acid is considered among the highly effective chemicals in stopping this reaction and preventing browning. It is a nontoxic and effective alternative to synthetic chemicals. It is also characterized by having powerful antioxidant and free radical scavenging properties. This review offers a focused and novel contribution to the scientific literature by exclusively investigating AA as an anti-browning agent (ABA), enabling a detailed understanding of its specific mechanisms, efficacy, and practical applications. Unlike broader reviews that cover many inhibitors, this work provides a comparative analysis of the performance of AA in various foods, highlighting its strengths and limitations in different contexts. By integrating research from past years, we highlight different approaches, such as the combination with synergists and integration with edible coatings and packaging. Importantly, we not only describe its optimal conditions and benefits, but also assess its limitations, such as its instability and susceptibility. Finally, the evidence and future directions are organized in a way that helps food technologists identify promising protocols, design preservation strategies, and avoid previously documented limitations of future research.

**Keywords**: Ascorbic acid, Browning control, Enzymatic browning, Fruits, Vegetables

### Introduction

Browning in fruits and vegetables (FV) is a major challenge to the food manufacturing industry due to the influence on appearance, nutritional value, and safety of the food product (Tinello & Lante, 2018). Consumers most often rely on visual cues to assess food quality and make informed decisions about freshness. The discoloration of the product contributes to decreased market viability and greater consumer rejection (Dias et al., 2020). In addition to the visual impact of discoloration, negatively browning also impacts utilization of produce and losses of production resources across the entire agricultural supply chain, from producers to retailers (Moon, Kwon, Lee, & Kim, 2020). Two major types of biochemical reactions cause this undesirable



discoloration: non-enzymatic browning and enzymatic browning. Non-enzymatic browning which is similar to the Maillard reaction, is more common in heat processing and less relevant for fresh produce. On the other hand, enzymatic browning (EB) dominates in fresh and minimally processed fruits and vegetables, where it causes tissue damage and rapid exposure to oxygen (Alrushdi, Mohammed et al., 2025; Paravisini & Peterson, 2019). The effects of EB on quality are particularly postharvest significant during handling, minimal processing. storage, and operations result in significant quality loss and economic cost (Moon et al., 2020). EB is associated with three enzymes: phenylalanine ammonia-lyase (PAL), peroxidase (POD) and polyphenol oxidase (PPO) (Arnold & Gramza-Michałowska, 2022). PPO (EC 1.10.3.1) catalyzes the EB reaction in plants, fungi, and bacteria. It is a copper-dependent cofactor protein, which can be activated as tyrosinase (monophenolase) catechol-oxidase or (diphenolase) (Sommano, Chanasut, Kumpoun, 2020).

This enzyme converts phenols to quinones, which combine with other substances to produce melanoid, a brown pigment (Lim & Wong, 2018; Singh et al., 2018). EB affects color retention and leads to nutritional and sensory losses, weakening marketability and reducing consumer acceptance. Therefore, it may be responsible for food waste (Shrestha et al., 2020). To mitigate the effects of EB, the fresh produce industry has traditionally relied on chemical inhibitors, but the rising concerns regarding the safety and regulatory status of synthetic additives, due to their possible health risks (e. g. allergic reactions), have increased focus on natural, food-safe alternatives (Bobo-García, Arroqui, Merino, & Vírseda, 2020; Dos Santos, López-Fernández, Favre. Mazzobre, & del Pilar Buera, Considering this, exploring novel natural additives for food without negative side effects that have anti-browning abilities is critically desirable for food-processors (Al-Abbasy, Ali, Rashan, & Al-Bajari, 2021). Ascorbic acid (AA), is a crucial natural substance found in abundance in plant sources. It can replace artificial additives to preserve FV, avoid costly losses, and extend shelf life. The use of AA in this field is due to its non-toxicity and lack of reported adverse effects in typical food applications (Dias et al., 2020; Paudel et al., 2020). AA has been selectively applied to various fresh foods (e.g, apples, potatoes, mushrooms, etc) and its potential use as a review preservative. This provides comprehensive summary by collecting and analyzing data exclusively on AA applications as an anti-browning agent in fresh products. Thus, it offers a scientific basis for food technologists to develop safe and effective antibrowning strategies.

### Search Criteria

Approximately 220 articles have been surveyed for this review. These articles were selected by a search process based on keywords such as ascorbic acid, browning, enzymatic browning, polyphenol oxidase, peroxidase, and browning control on electronic database systems such as PubMed, John Thompson, Scopus, Science Direct, and Google Scholar. After analyzing all the articles, many were excluded, and the research papers that were useful for this study were identified.

### **Browning**

The Browning phenomenon is widely considered a major problem facing the global economy. It is a biological process that often food and under different occurs in circumstances, which can have beneficial and harmful effects. For fruits, vegetables, and nuts, browning is considered undesirable. These foods lose their nutritional value and produce undesirable color, flavor, and taste. However, this process can be advantageous in processing other products such as chocolate, coffee, black tea (Debelo, Li, & Ferruzzi, 2020), bread, and soy sauce (Nooshkam, Varidi, & Bashash, 2019). It is necessary to create a distinctive color and flavor in these foods. The EB reaction (Fig. 1) is mainly caused by PPO (Arnold &

Gramza-Michałowska, 2022). PPO oxidizes diphenol to quinone, a highly sensitive active molecule subsequently polymerize into brown pigments by reacting with other phenolics, sugars, amino acids, or soluble proteins.

Fig. 1. The schematic of the enzymatic browning process (Moon et al., 2020) (modified)

It mostly happens in products during harvesting, storage, and processing (Queiroz, Mendes Lopes, Fialho, & Valente-Mesquita, 2008). The presence of EB degrades color, flavor, and nutritional content, resulting in reduced shelf life (Rashan & Al-abbasy, 2021). The rate of EB depends on the phenolic substrate contents, pH, oxygen accessibility, temperature, concentration, and source of an enzyme (Lante, Tinello, & Nicoletto, 2016).

### **Ascorbic Acid**

AA, commonly known as vitamin C, is a vital nutrient that can be acquired from various sources like fruits, vegetables, and multivitamin supplements (Ali et al., 2024; Murererehe, Uwitonze, Nikuze, Patel, & Razzaque, 2022). It plays a crucial role in numerous physiological functions within human health. AA is a sixcarbon lactone naturally produced from glucose and other sugars (Kükürt & Gelen, 2024; Mittu et al., 2022). Chemically, AA is a weak organic acid with an enediol structure conjugated with the carbonyl group in the lactone ring (Alburgus, Ali, & Al-abbasy, 2024; Atta, & Abdelgawad, Mohamed, 2017). compound's acidity and antioxidant properties are attributed to the two enolic hydrogen atoms supplying the electrons. Dehydroascorbic acid, which has the same vitamin activity as AA, is created when oxygen is present (Fig. 2) (Li, Guo, & Wang, 2008; Rico, Martin-Diana, Barat, & Barry-Ryan, 2007).

Fig. 2. Degradation of L-ascorbic acid to dehydroascorbic acid (Yin et al., 2022)

It is a natural component and broadly spread in FV (Kabasakalis, Siopidou, & Moshatou, 2000; Pernice et al., 2009), such as citrus fruits, strawberries, potatoes, tomato, broccoli, gooseberry, cabbage, turnip, red peppers, green peppers, and many green veggies. So, plants are becoming important sources of vitamin C (Malo & Wilson, 2000). It plays a role in numerous mono and dioxygenases to conserve

metals in a reduced state. For example, mono and dioxygenases commonly comprise copper or iron as redox cofactors. The food sector uses AA extensively because of its conservatory benefits, namely as the most potent and nontoxic antioxidant and a free-radical scavenger (Santos et al., 2022; Tu, Njus, & Schlegel, 2017). Since humans can not synthesize AA, it must be obtained through diet; it is considered

an important nutrient. However, during food storage, AA is quickly oxidized, which causes considerable losses (Hassan, Haddad, & Sultan, 2020). It is also acknowledged as the most important hydrophilic antioxidant for different types of enzymatic reactions (Bratovcic, 2020; Telang, 2013). This atypical antioxidant produces monodehydro ascorbate, a radical that combines preferentially with radicals rather than non-radical substances, and it gives a single reducing equivalent. This occurs because monodehydro ascorbate would form an energetically undesirable tricarbonyl structure if an electron were removed. AA oxidizes exclusively to monodehydroascorbate rather than producing this structure, and when monodehydroascorbate combines with other radicals, it oxidizes through processes that might prevent the creation of this undesirable structure (Du, Cullen, & Buettner, 2012). Reactive nitrogen and oxygen species, hydroperoxyl including radicals and superoxide, aqueous peroxyl radicals, and singlet oxygen, are easily scavenged by AA. (Yates, Schlicker, & Suitor, 1998).

### **Browning Control**

Browning control has always been a challenge for the food industry. Since phenols, oxygen, and enzymes are the three primary components contributing to browning processes, the primary goal of anti-browning agent treatments is to eliminate one of these variables and thus control browning (Younus, Mahdi, Al-Abbasy, & Sheej Ahmad, 2025). This control will extend the shelf life, improve customer acceptance of the food, and reduce economic losses (Bobo-García et al., 2020). To attenuate this undesirable process, industries employ chemical, physical, or biological techniques and treatments (Feng, Liu, Liu, Shi, & Wang, 2020; Zhang, He, & Simpson, 2018). These techniques have drawbacks, including changes in the sensory and nutritional qualities of the final products and potential health risks (Levaj et al., 2023). An efficient and widely used method for managing the EB of various fresh-cut FV is chemical treatment with antibrowning agents. These agents can be classified based on their mechanisms of action into several categories as chelating agents, reducing agents, acidulants, complexing agents, and enzyme inhibitors (Altunkaya & Gökmen, 2008, 2009). Successful browning inhibition by reducing agents, such as AA, erythorbic acid, ascorbyl-2-phosphate, ascorbyl-triphosphate, and cystein was observed (Farouk *et al.*, 2020; Sikora & Świeca, 2018).

# Anti-browning Mechanism of AA

Color is the key factor in appraising the consumer's quality of FV. Many natural and synthetic chemical compounds are commonly used to obstruct the browning rate of fresh FV through various mechanisms (Rodrigues, Asquieri, & Orsi, 2014). This treatment thus provides temporary protection to preserve these products from damage (Eleni & Theodoros, 2011). One of these important and extensively used compounds for this purpose is AA. Due to its safety and nutritional value, it is frequently employed as an anti-browning agent in dipping solutions (Pizzocaro, Torreggiani, & Gilardi, 1993; Veltman, Kho, Van Schaik, Sanders, & Oosterhaven, 2000). It has been investigated both alone and in conjunction with other antioxidants or preservatives, such as some phenols that function as competitive inhibitors like resorcinols (Yoruk & Marshall, 2003). This status is generally recognized as safe (GRAS). The Thailand Food and Drug Administration claims that there hasn't been any restriction for the upper limit of AA consumption (Suttirak & Manurakchinakorn, 2010). Understanding the kinetic and mechanistic pathways through which AA inhibits enzymatic browning is crucial for optimizing its application in food preservation. Kinetics studies on the rate of PPO inhibition by AA have shown that AA interacts with the enzyme's active site, preventing the oxidation of phenolic compounds (Ndakidemi, Mneney, Ndakidemi, 2014; Yoruk & Marshall, 2003). inhibitory action is The concentrationdependent, with higher AA concentrations leading to more significant inhibition of PPO

activity. Several studies have focused on elucidating these mechanisms:

- 1. Reduction of o-quinones back to phenolic substrates: As a strong reducing agent, AA converts the reactive o-quinones, the intermediate compounds produced during the PPO, back into their original phenolic forms before they can polymerize. This reaction prevents the accumulation of pigmented compounds and effectively halts browning at an early stage (Lee & Whitaker, 1995; Tang, Guo, & Zhu, 2023).

  2. Reduction of Cu<sup>2+</sup> to mononuclear copper
- Cu+ at the PPO active site (Suttirak & Manurakchinakorn, 2010).
- 3. Consumption of oxygen: AA can also react non-enzymatically with O2, reducing the oxygen availability required for PPO activity. This secondary mechanism further slows down quinone production and subsequently browning progression (Sapers & Miller, 1998).
- 4. AA acts as a competitive inhibitor of PPO by binding to the active center of the enzyme and competing with its natural substrates (Gao, Li, & Li, 2013).

On the other hand, AA may decrease the pH of crushed food products and thus inhibits the activities of POD and PPO or activates phenylpropanoid metabolism to raise total Degl'Innocenti, phenolic content (Landi, Guglielminetti, & Guidi, 2013; Li, Ding, Li, & Yan, 2023; Zhou et al., 2021). The efficiency of AA derivatives, containing dehydroascorbic acid, ascorbic acid-2-phosphate, isoascorbic acid-2-sulfate. acid. and ascorbic associated with that of AA (Hsu, Shieh, Bills, & White, 1988). Based on kinetic studies, AA and isoascorbic acid are the utmost effective inhibitors of mushroom PPO, followed by dehydroascorbic acid. According to (Özoğlu & Bayındırlı, 2002), AA is more effective than its isomer isoascorbic acid.

# Essential Motives for Using AA as an Anti**browning Agent**

Brown pigment on food products decreases consumer tolerability and affects their taste and nutritional status. Healthcare awareness has led to an increased focus on finding solutions to food browning (Singh et al., 2018). Consumers evaluate food products based on color, appearance, taste, and nutritional value (Peanparkdee & Iwamoto, 2019). Chemicalbased techniques were employed to prevent FV from browning. However, the application of this technique may pose health risks because some chemicals may include hazardous residues (Moon et al., 2020). Therefore, researchers resorted to using AA as an effective inhibitor of various PPOs (Arslan, Erzengin, Sinan, & Ozensoy, 2004; Chazarra, García-Carmona, & Cabanes, 2001). AA is a natural, abundantly available, and non-toxic material. Because of its great browning-inhibiting efficiency and the lack of deleterious effects in the foods to which it is introduced, AA is used as a reference or comparison to natural and synthetic chemicals, as well as natural extracts.

# **Effect of AA on Individual Commodities**

AA has been widely studied for its antioxidant effect against enzymatic browning in fruits and vegetables. This process is important for the maintenance of quality, color, and nutrient value of the commodity. AA efficacy on different commodities varies significantly with commodity type, processing type, and concentration of AA used. Literature has proven the effectiveness of AA in affecting the quality of apples and preventing fresh apples from turning brown. Granny Smith was less susceptible to browning upon immersion in (3% w/v) compared to other types (Yi et al., 2017). Similarly, another study on fresh apples pumila) concluded that PPO was (Malus inhibited by a large amount of AA (Wen et al., 2021). The impact of added AA stored at 4C° was investigated on POD and PPO reaction in light pretreated watermelon (Cucumis melo L. reticulatus Naud), and POD activity declined with time (And & Watson, 2001). The inhibitory efficacy of potatoes' PPO by chili pepper and lemon extracts was 70% and 54%, respectively. Anti-browning activity could be attributed to its high content of AA (Mercimek et al., 2015). In addition, the AA application to freshly harvested potatoes can postpone browning by inhibiting PAL (Zhou et al., 2021). AA is a potent competitive inhibitor of PPO's active site with catechol as a substrate to reduce the intensity of browning. This has been demonstrated in certain plants such as

lettuce (*Lactuca sativa*) (Altunkaya & Gökmen, 2008), Peppermint (*Mentha piperita*) (Kavrayan & Aydemir, 2001), and tea leaves (*Camellia sinensis*) (Öztürk, Aksoy, & Küfrevioğlu, 2020). Table 1 summarizes the inhibition of PPO activity in different vegetables and fruits using AA.

Table 1- Inhibitory effect of AA treatments on PPO activity in fruits and vegetables (\*(w/v) and \*\*mmol/L)

(*(w/v) and **mmol/L)				
Fruit/Vegetable	AA % PPO Concentration		References	
A 1. '	0.1760/ \$	inhibition	(C A 'C W' 1 ' 0 M 2022)	
Abiu	0.176% *	80%	(Susanto, Arif, Widayanti, & Matra, 2023)	
Apple (Red Delicious)	0.4 *	45%	(Arnold & Gramza-Michałowska, 2022)	
Apple puree	0.35% *	100	(Vercammen et al., 2012)	
Apple slices (Golden Delicious)	1% *	~25%	(Watada, Izumi, Luo, & Rodov, 2005)	
Apple slices (Newton Pippin)	1% *	~59%	(Watada et al., 2005)	
Apricot	2 **	100%	(Derardja, Pretzler, Kampatsikas, Barkat, & Rompel, 2019)	
Apricot	10 **	99%	(Morsy & Rayan, 2019)	
Banana	1% *	~85%	(Danyen, Boodia, & Ruggoo, 2009)	
Banana	0.5% *	~60%	(Moline, Buta, & Newman, 1999)	
Blackberry	0.1 **	100%	(Schwartz, Olgin, & Klinman, 2001)	
Blueberry	Varied	99.7%	(Siddiq & Dolan, 2017)	
Eggplant	0.5-1% *	15-25%	(Barbagallo, Chisarib, & Patanèc, 2012)	
Guava puree	1% *	~72%	(Khedr & Ali, 2017)	
Lettuce	5 **	~90%	(Landi et al., 2013)	
Lotus Rhizome	0.08**	86.21%	(Li, Li, Yan, & Wang, 2022)	
Mango	0.1% *	60%	(Arafat, 2009)	
Mango	1%*	75%	(Prasad, Sharma, & Srivastav, 2016)	
Mango	2% *	85-90%	(Marín <i>et al.</i> , 2021)	
Mango Puree	Varied	91.1% - 95.5%	(Guerrero-Beltrán, Swanson, & Barbosa-Cánovas, 2005)	
Mung Bean	2-20**	51–60%	(Sikora & Świeca, 2018)	
Mushroom	0.04 **	50%	(Hsu <i>et al.</i> , 1988)	
Peach	1**	95.4%	(Jia, Jiang, Chen, Wei, & Shao, 2022)	
Pear slices	1%*	75%	(Kader, Zagory, Kerbel, & Wang, 1989)	
Pineapple	0.05**	70%	(Auon-Reyna <i>et al.</i> , 2019)	
Pineapple slices	0.8% *	~50%	(Pérez-Gago, González-Aguilar, & Olivas, 2010)	
Pomegranate	150 **	100%	(Patil, Dhande, Thigale, & Rajput, 2011)	
Potato	1%*	65%	(Li et al., 2023)	
Potato slices	0.5%*	~65%	(AL-abbasy <i>et al.</i> , 2025)	
Strawberry	2% *	~88%	(Nazoori, Poraziz, Mirdehghan, Esmailizadeh, & Zamani Bahramabadi, 2020)	

### **Synergistic Treatments**

There have been various reports on the AA combination with other factors having a synergistic effect in inhibiting the action of the PPO and thus enhancing the control of the

browning of fresh foods. These synergists may be plant extracts, chemical compounds, or physical treatments. For instance, a synergistic influence of AA (0.05% w/v) and cinnamon essential oil nanoemulsion (0.1–0.4% w/v) was

found to completely inactivate PPO at 4 °C after two days of storage (Xu et al., 2020). AA with sodium dihydrogen phosphate controls the browning of yellow noodles by slightly reducing the PPO activity but significantly decreasing the free quinone content (Tang et al., 2023). The mango fruits were immersed in nanoparticles with calcium different concentrations of AA and stored for 35 days at 6 °C. The browning in kiwifruit segments can be effectively prevented by adding 0.3 g/L polyvinylpyrrolidone to the medium, which is followed by 0.2 mg/L AA. A mixture of 1.2% yerba mate leaf extract and 1% AA was proven to optimally suppress EB in fresh-cut Granny Smith apples (Rodríguez-Arzuaga Piagentini, 2018). Another study indicated that Rosa roxburghii extract, rich in AA and coumaric acid, inhibits the activity of PPO in Red Fuji cloudy juice (Yu et al., 2021). According to (Abdelwahd, Hakam, Labhilili, & Udupa, 2008; Babaei, Abdullah, Saleh, & Abdullah, 2013), the most effective method for managing EB was to pre-treat the tip of Faba bean seeds and Curculigo latifolia using polyvinylpyrrolidone and AA for nine hours. The PPO and POD activities responsible for browning of sugarcane juice were entirely inhibited when AA was combined with bentonite (Hithamani, Medappa, Chakkaravarthi, Ramalakshmi, & Raghavarao, 2018). AA and its isomer, erythorbic acid, were found to be equally effective in preventing browning in pineapple slices. The EB inhibition in apricots was investigated by inactivating their PPO using plant proteases and AA. Apricot PPO activity significantly declined to 66 and 55 % when treated with (0.1 and 0.2 mM mmol/L) of AA, respectively, compared to the untreated samples (Derardja et al., 2019). A

combination of AA and citric acid has been shown to significantly reduce the browning of fresh fruits such as Chinese Yam (Dioscorea polystachya Turczaninow). Citric acid can lower the pH of food products, hence increasing the efficacy of AA (Yang, Song, Wang, Wang, & Zhao, 2024). Alternatively, the combination of AA with chitosan (a natural polymer derived from chitin) has increased the ability to inhibit browning of plums and fresh-cut Chitosan acts as a protective environmental barrier, allowing AA to work more effectively, protecting the food from external air (Liu, Yuan, Chen, Li, & Liu, 2014; Özdemir & Gökmen, 2019). The ultrasound and AA showed a synergistic inhibitory effect on fresh-cut apple PPO activity during storage (Jang & Moon, 2011). The combination of AA (2g/L) with modified atmosphere packing inhibited the PPO activity and browning intensity of fresh-cut eggplant treated for four days (Sarengaowa et al., 2022). (Almeida & Nogueira, 1995) reported that PPO activity in banana, pear, and peach fruits was controlled using AA and heat treatment at 70°C for 2 minutes. The researchers highlighted the applicability of this method to substitute for the use of sulfur dioxide to control PPO activity.

### **Application in Juices**

The juice industry faces significant challenges with EB during the storage of fresh fruit juices. AA is one of the most extensively used natural preservatives to combat this issue. It has been shown through many studies that AA is effective in preventing this phenomenon that occurs in fruit juices. These effects are summarized in Table 2.

Table 2- Inhibitory effect of AA on different fruit juices

Fruit juice	AA concentration or combination	References
Banana	(1000 - 1200  ppm)	(Tapre & Jain, 2016)
Banana	(470  mg/L)	(Koffi, Sims, & Bates, 1991)
Apple	(1.8 mmol/L)	(Hope, 1961)
Apple	ascorbic and cyclodextrins	(Hicks, Sapers, & Seib, 1990)
Apple	ascorbic acid and acidic polyphosphate	(Sapers <i>et al.</i> , 1989)
Apple	0.2% w/v	(Komthong, Igura, & Shimoda, 2007)

Apple	-	(Ribárszki et al., 2022)
Cloudy apple	-	(Özoğlu & Bayındırlı, 2002)
Pear	-	(Montogomery & Petropakis, 1980)
Pear	0.24% (w/v)	(Jiang, Kim, Nam, Yim, & Eun, 2016)

# Common Trends, Optimal Conditions, and Knowledge Gaps

In compiling recent applications and developments of AA preservation, certain trends are common. First among these is the strong focus on multifunctional natural preservation techniques that accommodate consumer demand for sustainable and clean solutions. Synergistic mixtures and their use in edible or active packaging systems reflect a broader trend towards comprehensive antioxidant protection, combining physical barriers and biochemical activity to maintain freshness and prevent spoilage (Chua, Chan, Tay, & Soo, 2023; Oliveira et al., 2025). Optimum conditions for the efficacy of AA vary across commodities, but certain factors are universal in their implication. The concentration of AA tends to be in the range of 0.1-1%, being more potent for the browning controlling (Rasane et al., 2024). The pH of the processed product is a crucial factor, with the optimum value being slightly acidic (4-6), which is the most suitable environment for AA (Nicolas, Richard-Forget, Goupy, action Amiot, & Aubert, 1994). Moreover, the exposure to the light and temperature may affect AA stability, with studies showing that it is most effective when used at low storage temperature (0-10°C), thereby inhibiting rapid degradation and maintaining its anti-browning activity (Oms-Oliu et al., 2010, Paull, 1999). However, gaps in knowledge must be filled by conducting further research on long-term stability under conditions of fluctuating storage, compatibility with other natural preservatives, and scalability of packaging and packaging technologies. Although genetic approaches to increasing endogenous AA levels in crops hold much promise. However, regulatory concerns and public acceptability remain significant barriers (Oms-Oliu *et al.*, 2010; Paull, 1999).

# A Comparative Study

AA is widely recognized as one of the most effective natural anti-browning agents due to its strong reducing ability and non-toxicity. its effectiveness varies when However, compared to other chemical and natural inhibitors (Table 3), particularly in terms of mechanism of action, stability, cost, and sensory impact (Kim et al., 2019) . For example, citric acid acts as a metal chelator in preventing EB, through binding to the copper in PPO or pH-lowering, as mentioned by (Altunkaya & Gökmen, 2008). This applies to most carboxylic acids. However, the amino acid L-cysteine can inhibit the browning process principally by reducing POD and PPO activities via forming a stable complex with copper (Li et al., 2023; Sadoon & Ahmad, 2020).

Toxicologically, certain compounds, such as sulfites, especially in some sensitive persons, like asthmatics, are recognized to cause adverse health effects. Among this extremely sensitive group, many deaths have occurred as a result of consuming sulfur-containing foods. On the other hand, sulfates can also release SO2 gas in some foods, which harms the flavor of the product being treated (Taylor et al., 1988). These effects were not observed when using AA. Natural extracts are highly concentrationdependent in inhibiting enzymatic browning, which limits their use due to their strong taste or color (Gómez-López, 2002). In contrast, ascorbic acid is used at lower concentrations, is colorless and tasteless, and is consistently effective in a wide range of products (Rico et al., 2007).

Table 3- Com	narison of	anti-brov	vning agents	with	ascorbic acid
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Table 3- Comparison of anti-browning agents with ascorbic acid					
ABA	Mechanism of action	Effectiveness	Side effects/ Limitations		
Ascorbic acid	Reduces quinones back to phenols, acts as an antioxidant, and an oxygen scavenger	High short-term effectiveness, often combined with other agents for stability	Unstable, rapidly oxidizes, leading to browning rebound (re-browning)		
Citric acid	Lowers pH, reducing PPO activity, and chelates copper at the active site of PPO	Moderate, best when combined with other agents	Alters flavor due to acidity, limited by pH tolerance of the product		
Sodium metabisulfite	Reduces quinones, a strong PPO inhibitor, and reacts with o-quinones to form colorless products	Very high	Can cause allergic reactions (sulfite sensitivity), banned or limited in some countries		
Calcium ascorbate	Combining ascorbic acid's reduction with calcium's firming effect	Better than pure ascorbic acid in maintaining texture and color	Costly, limited by calcium-induced hardness in some products		
EDTA	Chelates copper ions from PPO, inhibiting enzyme activity	Moderate	Not approved for all food uses; may require labeling, synthetic additives		
Hexylresorcinol	Directly inhibits PPO; prevents formation of melanins	Very high	More expensive, limited regulatory approval in some regions		
Honey	Natural antioxidant, mild antimicrobial, and PPO inhibition	Low to moderate	May impart unwanted flavor/sweetness, variable composition		
L- Cysteine	Reacts with o-quinones to form colorless products; an antioxidant	High	May cause off-odors (sulfurous), allergen concerns		
Acetylated monoglycerides	Form coatings that limit oxygen contact, a physical barrier	Moderate (indirect)	Less effective alone, usually used with other agents		

### Future Trends in the Use of Ascorbic Acid as an Anti-**Browning Agent**

Changes in technology have expanded the potential uses of AA in food preservation, particularly when combined with other antioxidants or antimicrobials to produce efficacy with enhanced blends (Baghi, Gharsallaoui, Dumas, & Ghnimi, 2022). To increase its stability and extend its functional life, packaging systems such as nanoemulsions, biopolymer coatings, edible films, biodegradable coatings are being developed. This feature has oxygen-binding capabilities and provides continuous antioxidant protection, enhancing food quality and sustainability (Aziz, Ali, Haddad, & Abdullah, 2024; Kuai et al., 2021). On the processing front, treatments such as ultrasound-assisted dipping, vacuum impregnation, and spraying are being optimized to improve AA delivery and minimize waste (Díaz-Montes & Castro-Muñoz, 2021). In the long term, plant breeding and genetic

engineering methods seek to improve the natural growth or retention of AA in crops, reducing the need for external additives (Zhu, Wu, & Sun, 2019). Finally, with growing consumer demand for organic products with clean labels, AA's natural and safe properties are gaining popularity, and future applications are expected to focus on regulatory compliance and clear labeling.

### Limitation

AA is also frequently used in food applications but faces limitations due to instability and high susceptibility to oxidation, especially when exposed to air, and temperature (Comunian, Abbaspourrad, Favaro-Trindade, & Weitz, 2014; Wagner & Buettner, 2023). It may reduce its effectiveness over time and cause temporary effect. It can affect flavor and pH to become sour or tart, which is undesirable in fruits with delicate or low-acid flavors. This change in sensory

requires properties maximum dosage adjustment to be made carefully, especially in sensitive juice products where flavor balance is of the highest priority. These limitations underscore the importance of considering AA's chemical stability and sensory effect in welldesigned systems (Taqi, AL-shahery, & Al-Abbasy, 2024; Yin et al., 2022).

### Conclusion

The global food consumption landscape is rapidly transforming. Consumer preferences are driving a swift evolution in food choices worldwide. Color is one of the most important influencing consumer purchasing decisions. However, repeat purchases depend on the consumer's satisfaction with the product's texture, flavor, and nutritional value. EB is one of the serious problems to the food industry and represents the second most vital reason for the negative impact on color, which leads to deterioration. EB's progress has been related to PPO activity. Therefore, inhibiting certainly enzyme will limit development of this phenomenon. Applying anti-browning agents is considered the best effective way to control the EB reaction in fresh FV. In this regard, it is preferable to have natural agents such as AA. The widespread use of AA is due to its generally abundant, nontoxic, safe, and healthy properties compared to other chemicals. The capability of AA to prevent EB is represented by: reducing oquinones to diphenols or Cu<sup>2+</sup> to Cu+ at the PPO active site, while also decreasing crushed food pH. Based on the research considered, we conclude that AA was a good inhibitor for PPO and could prevent EB of FV. The anti-browning

efficiency of AA is related to two main factors: the types and cultivars of produce, and its concentration when it is alone or the concentrations of its synergetic factors. But often elevated concentrations had a greater effect than low ones. Also, the low pH at which AA works has a better effect in preventing the formation of brown pigmentation. After conducting this review and evaluating the general nutritional situation, we recommend using AA to prevent this phenomenon from forming on its own without combining it with other substances. This is to avoid the side effects of the combined substances on human health after consuming food products.

### **Funding Sources**

This research had no external funding.

### **Authors Contirbution**

**O.Y.A:** Conceptualization, Writing—review and editing, Supervision. Z.H.A: Data curation, Writing- original draft, Investigation. N.M.M: Formal analysisn, Writing-original draft. A.S.A: Validation, Data analysis, Software.

### Acknowledgments

The authors are grateful to the University of Mosul, which has provided enormous facilities and support to complete this review.

## **Abbreviation**

Anti-browning agent (ABA); Ascorbic acid (AA); Enzymatic browning (EB); fruits and Peroxidase vegetables (FV); (POD); Phenylalanine ammonia-lyase (PAL), Polyphenol oxidase (PPO).

### References

- 1. Abdelwahd, R., Hakam, N., Labhilili, M., & Udupa, S.M. (2008). Use of an adsorbent and antioxidants to reduce the effects of leached phenolics in in vitro plantlet regeneration of faba bean. African Journal of Biotechnology, 7(8). https://hdl.handle.net/20.500.11766/13068
- 2. Al-Abbasy, O.Y., Ali, W.I., Rashan, A.I., & Al-Bajari, S.A. (2021). Purification, characterization, and inhibition of tyrosinase from jerusalem artichoke (Helianthus tuberosus L.) tuber. Reports of Biochemistry & Molecular Biology, 10(3), 495. https://doi.org/10.52547/rbmb.10.3.495

- 3. AL-abbasy, O.Y., M Mahdi, N., Younus, S.A., Sheej Ahmad, O.A., & Al-Azzawi, A.G.S. (2025). Potato enzymatic browning, mitigation and prevention: Current overview of approaches and findings. Journal of Food Science and Technology (Iran), 22(164), 1-15.https://doi.org/10.22034/fsct.22.164.1
- 4. Al-burgus, A.F., Ali, A., & Al-abbasy, O.Y. (2024). New spiro-heterocyclic coumarin derivatives as antibacterial agents: design, synthesis and molecular docking. Chimica Techno Acta, 11(3). https://doi.org/10.15826/chimtech.2024.11.3.08
- 5. Ali, A., Riaz, S., Khalid, W., Fatima, M., Mubeen, U., Babar, Q., Manzoor, M.F., Zubair Khalid, M., & Madilo, F.K. (2024). Potential of ascorbic acid in human health against different diseases: an updated narrative review. International Journal of Food Properties, 27(1), 493-515. https://doi.org/10.1080/10942912.2024.2327335
- 6. Almeida, M., & Nogueira, J. (1995). The control of polyphenol oxidase activity in fruits and vegetables: a study of the interactions between the chemical compounds used and heat treatment. Plant Foods for Human Nutrition, 47, 245-256. https://doi.org/10.1007/BF01088333
- 7. Alrushdi, F.M.M., Mohammed Al-Abaasy, O.Y., Al-Saffar, R.N., Abbood, H.Y., Al-Hamairy, A.K., Saleh, M.Y., Abdelzaher, H., Abdelzaher, M.A., & A Kenawy, M. (2025). In vitro: inhibition of partially purified pancreatic ovine lipase by willow bark extracts. Journal of Bioscience and Applied Research, 11(1), 168-179. https://doi.org/10.21608/jbaar.2025.339535.1113
- 8. Altunkaya, A., & Gökmen, V. (2008). Effect of various inhibitors on enzymatic browning, antioxidant activity and total phenol content of fresh lettuce (Lactuca sativa). Food Chemistry, 107(3), 1173-1179. https://doi.org/10.1016/j.foodchem.2007.09.046
- 9. Altunkaya, A., & Gökmen, V. (2009). Effect of various anti-browning agents on phenolic compounds profile fresh lettuce (L.sativa). Food Chemistry, *117*(1), 122-126. https://doi.org/10.1016/j.foodchem.2009.03.085
- 10. And, O.L., & Watson, M. (2001). Effects of ascorbic acid on peroxidase and polyphenoloxidase activities in fresh-cut cantaloupe melon. Journal of Food Science, 66(9), 1283-1286. https://doi.org/10.1111/j.1365-2621.2001.tb15202.x
- 11. Arafat, L. (2009). Ascorbic acid and tissue browning in mango CV Hindi Be-Sennara fruits (Mangifera indica L.) under cold storage. Journal of Plant Production, 34(12), 11301-11310. https://doi.org/10.21608/jpp.2009.119187
- 12. Arnold, M., & Gramza-Michałowska, A. (2022). Enzymatic browning in apple products and its inhibition treatments: A comprehensive review. Comprehensive Reviews in Food Science and Food Safety, 21(6), 5038-5076. https://doi.org/10.1111/1541-4337.13059
- 13. Arslan, O., Erzengin, M., Sinan, S., & Ozensoy, O. (2004). Purification of mulberry (Morus alba L.) polyphenol oxidase by affinity chromatography and investigation of its kinetic and electrophoretic properties. Food Chemistry, 88(3), 479-484. https://doi.org/10.1016/j.foodchem.2004.04.005
- 14. Atta, E.M., Mohamed, N.H., & Abdelgawad, A.A. (2017). Antioxidants: An overview on the natural European Chemical and synthetic types. Bulletin 6(8), 365-375. https://doi.org/10.17628/ecb.2017.6.365-375
- 15. Ayon-Reyna, L.E., Ayon-Reyna, L.G., Lopez-Lopez, M.E., Lopez-Angulo, G., Pineda-Hidalgo, K.V., Zazuebla-Niebla, J.A., & Vega-Garcia, M.O. (2019). Changes in ascorbic acid and total phenolics contents associated with browning inhibition of pineapple slices. Food Science and Technology, 39, 531-537. https://doi.org/10.1590/fst.21117
- 16. Aziz, M.J., Ali, A.A., Haddad, M.F., & Abdullah, B.A. (2024). The relationship of vitamin D<sub>3</sub>, Ddimer, and antinuclear antibody levels with toxoplasmosis. Medical Journal of Babylon, 21(3), 556-559. https://doi.org/10.4103/mjbl.mjbl\_812\_23
- 17. Babaei, N., Abdullah, N.A.P., Saleh, G., & Abdullah, T.L. (2013). Control of contamination and explant browning in Curculigo latifolia in vitro cultures. Journal of Medicinal Plants Research, 7(8), 448-454. https://doi.org/10.5897/JMPR12.859
- 18. Baghi, F., Gharsallaoui, A., Dumas, E., & Ghnimi, S. (2022). Advancements in biodegradable active films for food packaging: Effects of nano/microcapsule incorporation. Foods, 11(5), 760. https://doi.org/10.3390/foods11050760

- 19. Barbagallo, R.N., Chisarib, M., & Patanèc, C. (2012). Use in vivo of natural anti-browning agents against polyphenol oxidase activity in minimally processed eggplant. Chemical Engineering, 27, 49-54. https://doi.org/10.3303/CET1227009
- 20. Bobo-García, G., Arroqui, C., Merino, G., & Vírseda, P. (2020). Antibrowning compounds for minimally processed potatoes: A review. Food Reviews International, 36(5), 529-546. https://doi.org/10.1080/87559129.2019.1650761
- 21. Bratovcic, A. (2020). Antioxidant enzymes and their role in preventing cell damage. Acta Scientific Nutritional Health, 4, 01-07. https://doi.org/10.31080/ASNH.2020.04.0659
- 22. Chazarra, S., García-Carmona, F., & Cabanes, J. (2001). Evidence for a tetrameric form of iceberg lettuce (Lactuca sativa L.) polyphenol oxidase: purification and characterization. Journal of Agricultural and Food Chemistry, 49(10), 4870-4875. https://doi.org/10.1021/jf0100301
- 23. Chua, L.S., Chan, Y.L., Tay, Z.Y., & Soo, J. (2023). Water-soluble propolis extract as a natural preservative for jaboticaba juice. Food Bioscience, 102651. https://doi.org/10.1016/j.fbio.2023.102651
- 24. Comunian, T.A., Abbaspourrad, A., Favaro-Trindade, C.S., & Weitz, D.A. (2014). Fabrication of solid lipid microcapsules containing ascorbic acid using a microfluidic technique. Food Chemistry, 152, 271-275. https://doi.org/10.1016/j.foodchem.2013.11.149
- 25. Danyen, S.B., Boodia, N., & Ruggoo, A. (2009). Interaction effects between ascorbic acid and calcium chloride in minimizing browning of fresh-cut green banana slices. Journal of Food Processing and Preservation, 33, 12-26. https://doi.org/10.1111/j.1745-4549.2008.00246.x
- 26. Debelo, H., Li, M., & Ferruzzi, M.G. (2020). Processing influences on food polyphenol profiles and biological activity. Current **Opinion** in Food Science, 32, 90-102. https://doi.org/10.1016/j.cofs.2020.03.001
- 27. Dias, C., Fonseca, A.M., Amaro, A.L., Vilas-Boas, A.A., Oliveira, A., Santos, S.A., Silvestre, A.J., Rocha, S.M., Isidoro, N., & Pintado, M. (2020). Natural-based antioxidant extracts as potential mitigators of fruit browning. Antioxidants, 9(8), 715. https://doi.org/10.3390/antiox9080715
- 28. Díaz-Montes, E., & Castro-Muñoz, R. (2021). Edible films and coatings as food-quality preservers: An overview. Foods, 10(2), 249. https://doi.org/10.3390/foods10020249
- 29. Du, J., Cullen, J.J., & Buettner, G.R. (2012). Ascorbic acid: chemistry, biology and the treatment of cancer. Biochimica et Biophysica Acta (BBA)-Reviews on Cancer, 1826(2), 443-457. https://doi.org/10.1016/j.bbcan.2012.06.003
- 30. eddine Derardja, A., Pretzler, M., Kampatsikas, I., Barkat, M., & Rompel, A. (2019). Inhibition of apricot polyphenol oxidase by combinations of plant proteases and ascorbic acid. Food Chemistry: X, 4, 100053. https://doi.org/10.1016/j.fochx.2019.100053
- 31. Eleni, M., & Theodoros, V. (2011). Effect of storage conditions on the sensory quality, colour and texture of fresh-cut minimally processed cabbage with the addition of ascorbic acid, citric acid and calcium chloride. Food and Nutrition Sciences, 2011. https://doi.org/10.4236/fns.2011.29130
- 32. Farouk, B., Aref, N., Rachid, C., Mourad, L., Emna, K., Fethi, B., Rania, B., Wafa, N., Kenza, B., & Boumediene, M. (2020). Characterization of three polyphenol oxidase isoforms in royal dates and inhibition of its enzymatic browning reaction by indole-3-acetic acid. International Journal of Biological Macromolecules, 145, 894-903. https://doi.org/10.1016/j.ijbiomac.2019.09.140
- 33. Favre, L.C., Dos Santos, C., López-Fernández, M.P., Mazzobre, M.F., & del Pilar Buera, M. (2018). Optimization of \( \beta\)-cyclodextrin-based extraction of antioxidant and anti-browning activities from leaves by response surface methodology. FoodChemistry, 86-95. https://doi.org/10.1016/j.foodchem.2018.05.078
- 34. Feng, Y., Liu, Q., Liu, P., Shi, J., & Wang, Q. (2020). Aspartic acid can effectively prevent the enzymatic browning of potato by regulating the generation and transformation of brown product. Biology Technology, 111209. **Postharvest** and 166, https://doi.org/10.1016/j.postharvbio.2020.111209

- 35. Gao, L., Li, K.D., & Li, Y.C. (2013). Inhibition kinetics of polyphenol oxidase from purple sweet ascorbic acid. Applied Mechanics and Materials, https://doi.org/10.4028/www.scientific.net/AMM.333-335.1921
- 36. Gómez-López, V.M. (2002). Some biochemical properties of polyphenol oxidase from two varieties of avocado. Food Chemistry, 77(2), 163-169. https://doi.org/10.1016/S0308-8146(01)00331-4
- 37. Guerrero-Beltrán, J.A., Swanson, B.G., & Barbosa-Cánovas, G.V. (2005). Inhibition of polyphenoloxidase in mango puree with 4-hexylresorcinol, cysteine and ascorbic acid. LWT-Food Science and Technology, 38(6), 625-630. https://doi.org/10.1016/j.lwt.2004.08.002
- 38. Hassan, M.N., Haddad, M.F., & Sultan, S.M. (2020). Inhibition of Staphylococcus aureus growth isolated from teeth decay using pomegranate fat extract fortified by silver nanoparticles (AgNp). (09752366),**International** Journal Pharmaceutical Research of 12(4). https://doi.org/10.31838/ijpr/2020.12.04.240
- 39. Hicks, K.B., Sapers, G.M., & Seib, P.A. (1990). Process for preserving raw fruit and vegetable juices using cyclodextrins and compositions thereof. In): Google Patents.
- 40. Hithamani, G., Medappa, H., Chakkaravarthi, A., Ramalakshmi, K., & Raghavarao, K.S.M.S. (2018). Effect of adsorbent and acidulants on enzymatic browning of sugarcane juice. Journal of Food Science and Technology, 55, 4356-4362. https://doi.org/10.1007/s13197-018-3350-4
- 41. Hope, G. (1961). Use of antioxidants in caning apple halves. Food Technology, 15(12), 548-560.
- 42. Hsu, A., Shieh, J., Bills, D., & White, K. (1988). Inhibition of mushroom polyphenoloxidase by ascorbic acid derivatives. Journal of Food Science, 53(3), 765-767. https://doi.org/10.1111/j.1365-2621.1988.tb08951.x
- 43. Jang, J.-H., & Moon, K.-D. (2011). Inhibition of polyphenol oxidase and peroxidase activities on fresh-cut apple by simultaneous treatment of ultrasound and ascorbic acid. Food Chemistry, 124(2), 444-449. https://doi.org/10.1016/j.foodchem.2010.06.052
- 44. Jia, S., Jiang, S., Chen, Y., Wei, Y., & Shao, X. (2022). Comparison of inhibitory effects of cinnamic acid, β-cyclodextrin, L-cysteine, and ascorbic acid on soluble and membrane-bound polyphenol oxidase in peach fruit. Foods, 12(1), 167. https://doi.org/10.3390/foods12010167
- 45. Jiang, G.-H., Kim, Y.-M., Nam, S.-H., Yim, S.-H., & Eun, J.-B. (2016). Enzymatic browning inhibition and antioxidant activity of pear juice from a new cultivar of asian pear (Pyrus pyrifolia Nakai cv. Sinhwa) with different concentrations of ascorbic acid. Food Science and Biotechnology, 25, 153-158. https://doi.org/10.1007/s10068-016-0023-9
- 46. Kabasakalis, V., Siopidou, D., & Moshatou, E. (2000). Ascorbic acid content of commercial fruit and its rate of loss upon storage. Food Chemistry, 70(3),325-328. https://doi.org/10.1016/S0308-8146(00)00093-5
- 47. Kader, A.A., Zagory, D., Kerbel, E.L., & Wang, C.Y. (1989). Modified atmosphere packaging of fruits vegetables. Critical Reviews inFoodScience & Nutrition, https://doi.org/10.1080/10408398909527490
- 48. Kavrayan, D., & Aydemir, T. (2001). Partial purification and characterization of polyphenoloxidase from peppermint (Mentha piperita). Food Chemistry, 74(2), 147-154. https://doi.org/10.1016/S0308-8146(01)00106-6
- 49. Khedr, E., & Ali, M. (2017). Application of antibrowning and firmness supporting compounds to maintain the quality of fresh-cut guava. Egyptian Journal of Agricultural Sciences, 68(3), 293-303. https://doi.org/10.21608/ejarc.2017.212733
- 50. Kim, T.-K., Hwang, K.-E., Lee, M.-A., Paik, H.-D., Kim, Y.-B., & Choi, Y.-S. (2019). Quality characteristics of pork loin cured with green nitrite source and some organic acids. *Meat Science*, 152, 141-145. https://doi.org/10.1016/j.meatsci.2019.02.015
- 51. Koffi, E., Sims, C., & Bates, R. (1991). Viscosity reduction and prevention of browning in the preparation of clarified banana juice 1. Journal of Food Quality, 14(3), 209-218. https://doi.org/10.1111/j.1745-4557.1991.tb00062.x
- 52. Komthong, P., Igura, N., & Shimoda, M. (2007). Effect of ascorbic acid on the odours of cloudy apple juice. Food Chemistry, 100(4), 1342-1349. https://doi.org/10.1016/j.foodchem.2005.10.070

- 53. Kuai, L., Liu, F., Chiou, B.-S., Avena-Bustillos, R.J., McHugh, T.H., & Zhong, F. (2021). Controlled release of antioxidants from active food packaging: A review. Food Hydrocolloids, 120, 106992. https://doi.org/10.1016/j.foodhyd.2021.106992
- 54. Kükürt. A.. & Gelen, V. (2024). Ascorbic Acid: Biochemistry (https://doi.org/10.5772/intechopen.105286): BoD–Books on Demand.
- 55. Landi, M., Degl'Innocenti, E., Guglielminetti, L., & Guidi, L. (2013). Role of ascorbic acid in the inhibition of polyphenol oxidase and the prevention of browning in different browning-sensitive Lactuca sativa var. capitata (L.) and Eruca sativa (Mill.) stored as fresh-cut produce. Journal of the Science of Food and Agriculture, 93(8), 1814-1819, https://doi.org/10.1002/jsfa.5969
- 56. Lante, A., Tinello, F., & Nicoletto, M. (2016). UV-A light treatment for controlling enzymatic browning of fresh-cut fruits. Innovative Food Science & Emerging Technologies, 34, 141-147. https://doi.org/10.1016/j.ifset.2015.12.029
- 57. Lee, C.Y., & Whitaker, J.R. (1995). Enzymatic browning and its prevention. ACS Publications. https://doi.org/10.1021/bk-1995-0600.ch001
- 58. Levaj, B., Pelaić, Z., Galić, K., Kurek, M., Ščetar, M., Poljak, M., Dite Hunjek, D., Pedisić, S., Balbino, S., & Čošić, Z. (2023). Maintaining the quality and safety of fresh-cut potatoes (Solanum tuberosum): Overview of recent findings and approaches. Agronomy, https://doi.org/10.3390/agronomy13082002
- 59. Li, C., Ding, X., Li, J., & Yan, S. (2023). Effects of different concentrations of ascorbic acid on the stability of (+)-Catechin under enzymatic conditions. Food Chemistry, 399, 133933. https://doi.org/10.1016/j.foodchem.2022.133933
- 60. Li, C., Li, J., Yan, S., & Wang, Q. (2022). The mechanism of interaction between lotus rhizome polyphenol oxidase and ascorbic acid: Inhibitory activity, thermodynamics, and conformation analysis. Journal of Food Biochemistry, 46(5), e14047. https://doi.org/10.1111/jfbc.14047
- 61. Li, G., Wang, X., Zhu, H., Li, G., Du, J., Song, X., & Erihemu. (2023). Use of different food additives to control browning in fresh-cut potatoes. Food Science & Nutrition, 11(12), 7967-7973. https://doi.org/10.1002/fsn3.3714
- 62. Li, H., Guo, A., & Wang, H. (2008). Mechanisms of oxidative browning of wine. Food Chemistry, 108(1), 1-13. https://doi.org/10.1016/j.foodchem.2007.10.065
- 63. Lim, W.Y., & Wong, C.W. (2018). Inhibitory effect of chemical and natural anti-browning agents on polyphenol oxidase from ginger (Zingiber officinale Roscoe). Journal of Food Science and Technology, 55, 3001-3007. https://doi.org/10.1007/s13197-018-3218-7
- 64. Liu, K., Yuan, C., Chen, Y., Li, H., & Liu, J. (2014). Combined effects of ascorbic acid and chitosan on the quality maintenance and shelf life of plums. Scientia Horticulturae, 176, 45-53. https://doi.org/10.1016/j.scienta.2014.06.027
- 65. Malo, C., & Wilson, J. (2000). Glucose modulates vitamin C transport in adult human small intestinal border brush membrane vesicles. The Journal Nutrition, *130*(1), of https://doi.org/10.1093/jn/130.1.63
- 66. Marín, A., Baldwin, E.A., Bai, J., Wood, D., Ference, C., Sun, X., Brecht, J. K., & Plotto, A. (2021). Edible coatings as carriers of antibrowning compounds to maintain appealing appearance of fresh-cut mango. HortTechnology, 31(1), 27-35. https://doi.org/10.21273/HORTTECH04687-20
- 67. Mercimek, H.A., Guzeldag, G., Ucan, F., Guler, K.C., Karaman, M., & Karavilan, R. (2015). Inhibition of polyphenol oxidase purified from potato (Solanum tuberosum). Romanian Biotechnological Letters, 20(6), 10961-10968. https://doi.org/10961-10968
- 68. Mittu, B., Bhat, Z.R., Chauhan, A., Kour, J., Behera, A., & Kaur, M. (2022). Ascorbic acid. In Nutraceuticals and Health Care, (pp. 289-302): Elsevier. https://doi.org/10.1016/b978-0-323-89779-2.00015-6
- 69. Moline, H., Buta, J., & Newman, I. (1999). Prevention of browning of banana slices using natural and their derivatives 1. Journal of Food Quality, 499-511. https://doi.org/10.1111/j.1745-4557.1999.tb00181.x

- 70. Montogomery, M., & Petropakis, H. (1980). Inactivation of Bartlett pear polyphenol oxidase with heat presence of ascorbic acid. Journal of Food Science, 45(4), https://doi.org/10.1111/j.1365-2621.1980.tb07529.x
- 71. Moon, K.M., Kwon, E.-B., Lee, B., & Kim, C.Y. (2020). Recent trends in controlling the enzymatic browning of fruit and vegetable products. Molecules, 25(12), 2754. https://doi.org/10.3390/molecules25122754
- 72. Morsy, N.E., & Rayan, A.M. (2019). Effect of different edible coatings on biochemical quality and shelf life of apricots (Prunus armenica L. cv Canino). Journal of Food Measurement and Characterization, 13, 3173-3182. https://doi.org/10.1007/s11694-019-00240-2
- 73. Murererehe, J., Uwitonze, A.M., Nikuze, P., Patel, J., & Razzaque, M.S. (2022). Beneficial effects of vitamin C in maintaining optimal oral health. Frontiers in Nutrition, 8, 805809. https://doi.org/10.3389/fnut.2021.805809
- 74. Nazoori, F., Poraziz, S., Mirdehghan, S.H., Esmailizadeh, M., & ZamaniBahramabadi, E. (2020). Improving shelf life of strawberry through application of sodium alginate and ascorbic acid coatings. Journal of Horticultural and Technology, International Science https://doi.org/10.22059/ijhst.2020.297134.341
- 75. Ndakidemi, C.F., Mneney, E., & Ndakidemi, P.A. (2014). Effects of ascorbic acid in controlling lethal browning in in vitro culture of Brahylaena huillensis using nodal segments. American Journal of Plant Sciences, 2014. https://doi.org/10.4236/ajps.2014.51024
- 76. Nicolas, J.J., Richard-Forget, F.C., Goupy, P.M., Amiot, M.J., & Aubert, S.Y. (1994). Enzymatic browning reactions in apple and apple products. Critical Reviews in Food Science & Nutrition, 34(2), 109-157. https://doi.org/10.1080/10408399409527653
- 77. Nooshkam, M., Varidi, M., & Bashash, M. (2019). The Maillard reaction products as food-born antioxidant and antibrowning agents in model and real food systems. Food Chemistry, 275, 644-660. https://doi.org/10.1016/j.foodchem.2018.09.083
- 78. Oliveira, I., Pinto, T., Afonso, S., Karaś, M., Szymanowska, U., Gonçalves, B., & Vilela, A. (2025). Sustainability in bio-based edible films, coatings, and packaging for small fruits. Applied Sciences, 15(3), 1462. https://doi.org/10.3390/app15031462
- 79. Oms-Oliu, G., Rojas-Graü, M.A., González, L.A., Varela, P., Soliva-Fortuny, R., Hernando, M.I.H., Munuera, I.P., Fiszman, S., & Martín-Belloso, O. (2010). Recent approaches using chemical treatments to preserve quality of fresh-cut fruit: A review. Postharvest Biology and Technology, 57(3), 139-148. https://doi.org/10.1016/j.postharvbio.2010.04.001
- 80. Özdemir, K.S., & Gökmen, V. (2019). Effect of chitosan-ascorbic acid coatings on the refrigerated storage stability of fresh-cut apples. Coatings, 9(8), 503. https://doi.org/10.3390/coatings9080503
- 81. Özoğlu, H., & Bayındırlı, A. (2002). Inhibition of enzymic browning in cloudy apple juice with selected antibrowning agents. Food Control, 13(4-5), 213-221. https://doi.org/10.1016/S0956-7135(02)00011-7
- 82. Öztürk, C., Aksoy, M., & Küfrevioğlu, Ö.İ. (2020). Purification of tea leaf (Camellia sinensis) polyphenol oxidase by using affinity chromatography and investigation of its kinetic properties. Journal of Food Measurement and Characterization, 14, 31-38.https://doi.org/10.1007/s11694-019-00264-8
- 83. Paravisini, L., & Peterson, D.G. (2019). Mechanisms non-enzymatic browning in orange juice during storage. Food Chemistry, 289, 320-327.https://doi.org/10.1016/j.foodchem.2019.03.049
- 84. Patil, V.M., Dhande, G., Thigale, D.M., & Rajput, J. (2011). Micropropagation of pomegranate (Punica granatum L.) Bhagava' cultivar from nodal explant. African Journal of Biotechnology, 10(79), 18130-18136. https://doi.org/10.5897/AJB11.1437
- 85. Paudel, P., Seong, S.H., Wagle, A., Min, B.S., Jung, H.A., & Choi, J.S. (2020). Antioxidant and antibrowning property of 2-arylbenzofuran derivatives from *Morus alba* Linn root bark. *Food Chemistry*, 309, 125739. https://doi.org/10.1016/j.foodchem.2019.125739
- 86. Paull, R. (1999). Effect of temperature and relative humidity on fresh commodity quality. *Postharvest* Biology and Technology, 15(3), 263-277. https://doi.org/10.1016/S0925-5214(98)00090-8

- 87. Peanparkdee, M., & Iwamoto, S. (2019). Bioactive compounds from by-products of rice cultivation and rice processing: Extraction and application in the food and pharmaceutical industries. Trends in Food Science & Technology, 86, 109-117. https://doi.org/10.1016/j.tifs.2019.02.041
- 88. Pérez-Gago, M.B., González-Aguilar, G., & Olivas, G. (2010). Edible coatings for fruits and vegetables. https://doi.org/10.2212/spr.2010.3.4
- 89. Pernice, R., Borriello, G., Ferracane, R., Borrelli, R.C., Cennamo, F., & Ritieni, A. (2009). Bergamot: A source of natural antioxidants for functionalized fruit juices. Food Chemistry, 112(3), 545-550. https://doi.org/10.1016/j.foodchem.2008.06.004
- 90. Pizzocaro, F., Torreggiani, D., & Gilardi, G. (1993). Inhibition of apple polyphenoloxidase (PPO) by ascorbic acid, citric acid and sodium chloride. Journal of Food Processing and Preservation, 17(1), 21-30. https://doi.org/10.1111/j.1745-4549.1993.tb00223.x
- 91. Prasad, K., Sharma, R., & Srivastav, M. (2016). Postharvest treatment of antioxidant reduces lenticel browning and improves cosmetic appeal of mango (Mangifera indica L.) fruits without impairing quality. Journal of Food Science and Technology, 53, 2995-3001. https://doi.org/10.1007/s13197-016-2267-z
- 92. Queiroz, C., Mendes Lopes, M.L., Fialho, E., & Valente-Mesquita, V.L. (2008). Polyphenol oxidase: characteristics and mechanisms of browning control. Food Reviews International, 24(4), 361-375. https://doi.org/10.1080/87559120802089332
- 93. Rasane, P., Singh, J., Kaur, S., Bakshi, M., Gunjal, M., Kaur, J., Sharma, K., Sachan, S., Singh, A., & Bhadariya, V. (2024). Strategic advances in the management of browning in fruits and vegetables. Food and Bioprocess Technology, 17(2), 325-350. https://doi.org/10.1007/s11947-023-03128-8
- 94. Rashan, A.I., & Al-abbasy, O.Y. (2021). Inhibitory and kinetic study of partially purified tyrosinase from Iraqi quince fruit. Plant Cell Biotechnology and Molecular Biology, 22(23-24), 1-14. https://hal.science/hal-05172822v1
- 95. Ribárszki, Á., Székely, D., Szabó-Nótin, B., Góczán, B., Friedrich, L., Nguyen, Q., & Máté, M. (2022). Effect of ascorbic acid and acerola juice on some quality properties of aseptic filled apple juice. Acta Alimentaria, https://doi.org/10.1556/066.2022.00030
- 96. Rico, D., Martin-Diana, A.B., Barat, J., & Barry-Ryan, C. (2007). Extending and measuring the quality of fresh-cut fruit and vegetables: a review. Trends in Food Science & Technology, 18(7), 373-386. https://doi.org/10.1016/j.tifs.2007.03.011
- 97. Rodrigues, O.R.L., Asquieri, E.R., & Orsi, D.C. (2014). Prevention of enzymatic browning of vacon flour by the combined use of anti-browning agents and the study of its chemical composition. Food Science and Technology, 34, 275-280. https://doi.org/10.1590/fst.2014.0045
- 98. Rodríguez-Arzuaga, M., & Piagentini, A.M. (2018). New antioxidant treatment with yerba mate (*Ilex* paraguariensis) infusion for fresh-cut apples: Modeling, optimization, and acceptability. Food Science and Technology International, 24(3), 223-231. https://doi.org/10.1177/1082013217744424
- 99. Sadoon, A.M., & Ahmad, O.S. (2020). Spectroscopy study of MCL<sub>2</sub>(H<sub>2</sub>O)N cluster using AB initio calculations. Periódico Química, 584-597. https://doi.org/10.52571/ptq.v17.n36.2020.599\_periodico36\_pgs\_584\_597.pdf
- Santos, K.L., Bragança, V.A., Pacheco, L.V., Ota, S.S., Aguiar, C.P., & Borges, R.S. (2022). 100. Essential features for antioxidant capacity of ascorbic acid (vitamin C). Journal of Molecular Modeling, 28, 1-8. https://doi.org/10.1007/s00894-021-04994-9
- 101. Sapers, G., Hicks, K., Phillips, J., Garzarella, L., Pondish, D., Matulaitis, R., McCormack, T., Sondey, S., Seib, P., & Ei-Atawy, Y. (1989). Control of enzymatic browning in apple with ascorbic acid derivatives, polyphenol oxidase inhibitors, and complexing agents. Journal of Food Science, 54(4), 997-1002. https://doi.org/10.1111/j.1365-2621.1989.tb07931.x
- Sapers, G.M., & Miller, R.L. (1998). Browning inhibition in fresh-cut pears. *Journal of Food* Science, 63(2), 342-346. https://doi.org/10.1111/j.1365-2621.1998.tb15738.x
- Sarengaowa, Wang, L., Liu, Y., Yang, C., Feng, K., & Hu, W. (2022). Effect of ascorbic acid 103. combined with modified atmosphere packaging for browning of fresh-cut eggplant. Coatings, 12(10), 1580. https://doi.org/10.3390/coatings12101580

- 104. Schwartz, B., Olgin, A.K., & Klinman, J.P. (2001). The role of copper in topa quinone biogenesis and catalysis, as probed by azide inhibition of a copper amine oxidase from yeast. Biochemistry, 40(9), 2954-2963. https://doi.org/10.1021/bi0021378
- 105. Shrestha, L., Kulig, B., Moscetti, R., Massantini, R., Pawelzik, E., Hensel, O., & Sturm, B. (2020). Optimisation of physical and chemical treatments to control browning development and enzymatic activity on fresh-cut apple slices. Foods, 9(1), 76. https://doi.org/10.3390/foods9010076
- Siddiq, M., & Dolan, K. (2017). Characterization of polyphenol oxidase from blueberry 106. (Vaccinium corymbosum L.). Food Chemistry, 216-220. https://doi.org/10.1016/j.foodchem.2016.09.061
- Sikora, M., & Świeca, M. (2018). Effect of ascorbic acid postharvest treatment on enzymatic 107. browning, phenolics and antioxidant capacity of stored mung bean sprouts. Food Chemistry, 239, 1160-1166. https://doi.org/10.1016/j.foodchem.2017.07.067
- 108. Singh, B., Suri, K., Shevkani, K., Kaur, A., Kaur, A., & Singh, N. (2018). Enzymatic browning of fruit and vegetables: A review. Enzymes in Food Technology: Improvements and Innovations, 63-78. https://doi.org/10.1007/978-981-13-1933-4 4
- 109. Sommano, S.R., Chanasut, U., & Kumpoun, W. (2020). Enzymatic browning and its amelioration in fresh-cut tropical fruits. In Fresh-cut Fruits and Vegetables, (pp. 51-76): Elsevier. https://doi.org/10.1016/b978-0-12-816184-5.00003-3
- 110. Susanto, S., Arif, A.B., Widayanti, S.M., & Matra, D.D. (2023). Ascorbic acid extends the shelf-life of Abiu (*Pouteria caimito*) fruit by maintaining quality and delaying browning symptoms. The Horticulture Journal, 92(3), 216-226. https://doi.org/10.2503/hortj.QH-053
- Suttirak, W., & Manurakchinakorn, S. (2010). Potential application of ascorbic acid, citric acid and oxalic acid for browning inhibition in fresh-cut fruits and vegetables. Walailak Journal of Science and Technology (WJST), 7(1), 5-14. https://doi.org/10.2004/wjst.v7i1.47
- Tang, Y.-Y., Guo, X.-N., & Zhu, K.-X. (2023). Inhibitory mechanism of sodium dihydrogen 112. phosphate and ascorbic acid on browning in yellow alkaline noodles. Journal of Cereal Science, 112, 103706. https://doi.org/10.1016/j.jcs.2023.103706
- 113. Tapre, A., & Jain, R. (2016). Study of inhibition of browning of clarified banana juice. Asian Journal of Dairy and Food Research, 35(2), 155-159. https://doi.org/10.18805/ajdfr.v35i2.10723
- 114. Taqi, H.M., AL-shahery, Y.J., & Al-Abbasy, O.Y. (2024). Innovative isolation of nostoc minutum protein for antibacterial applications. Egyptian Journal of Aquatic Biology and Fisheries, 28(6), 2055-2071. https://doi.org/10.21608/ejabf.2024.400573
- Taylor, S.L., Bush, R.K., Selner, J.C., Nordlee, J.A., Wiener, M.B., Holden, K., Koepke, J. 115. W., & Busse, W.W. (1988). Sensitivity to sulfited foods among sulfite-sensitive subjects with asthma. Journal of Allergy and Clinical Immunology, 81(6), 1159-1167. https://doi.org/10.1016/0091-6749(88)90885-8
- 116. Telang, P.S. (2013). Vitamin C in dermatology. *Indian Dermatology Online Journal*, 4(2), 143-146. https://doi.org/10.4103/2229-5178.110593
- Tinello, F., & Lante, A. (2018). Recent advances in controlling polyphenol oxidase activity of 117. fruit and vegetable products. Innovative Food Science & Emerging Technologies, 50, 73-83. https://doi.org/10.1016/j.ifset.2018.10.008
- Tu, Y.-J., Njus, D., & Schlegel, H.B. (2017). A theoretical study of ascorbic acid oxidation 118. and HOO/O 2- radical scavenging. Organic & Biomolecular Chemistry, 15(20), 4417-4431. https://doi.org/10.1039/C7OB00791D
- Veltman, R., Kho, R., Van Schaik, A., Sanders, M., & Oosterhaven, J. (2000). Ascorbic acid 119. and tissue browning in pears (Pyrus communis L. cvs Rocha and Conference) under controlled atmosphere conditions. **Postharvest** Biology and Technology, *19*(2), 129-137. https://doi.org/10.1016/S0925-5214(00)00095-8
- Vercammen, A., Vanoirbeek, K.G., Lemmens, L., Lurquin, I., Hendrickx, M.E., & Michiels, 120. C.W. (2012). High pressure pasteurization of apple pieces in syrup: microbiological shelf-life and

- quality evolution during refrigerated storage. Innovative Food Science & Emerging Technologies, 16, 259-266. https://doi.org/10.1016/j.ifset.2012.06.009
- Wagner, B.A., & Buettner, G.R. (2023). Stability of aqueous solutions of ascorbate for basic 121. research and for intravenous administration. Advances in Redox Research, 9, 100077. https://doi.org/10.1016/j.arres.2023.100077
- Watada, A.E., Izumi, H., Luo, Y., & Rodov, V. (2005). Fresh-cut produce. Environmentally 122. friendly technologies for agricultural produce quality, 149-203. https://doi.org/10.1201/9780203500361.ch7
- Wen, Y.T., Liang, Y.Q., Chai, W.M., Wei, Q.M., Yu, Z.Y., & Wang, L.J. (2021). Effect of ascorbic acid on tyrosinase and its anti-browning activity in fresh-cut Fuji apple. Journal of Food Biochemistry, 45(12), e13995. https://doi.org/10.1111/jfbc.13995
- Xu, J., Zhou, L., Miao, J., Yu, W., Zou, L., Zhou, W., Liu, C., & Liu, W. (2020). Effect of 124. cinnamon essential oil nanoemulsion combined with ascorbic acid on enzymatic browning of cloudy apple juice. Food and Bioprocess Technology, 13, 860-870. https://doi.org/10.1007/s11947-020-02443-8
- 125. Yang, W., Song, X., Wang, Q., Wang, W., & Zhao, Z. (2024). Combined addition of citric acid and ascorbic acid significantly inhibits browning in Chinese yam (Dioscorea polystachya Turczaninow) processing. **Journal** of Food Quality, 2024(1), 1197382. https://doi.org/10.1155/2024/1197382
- Yates, A.A., Schlicker, S.A., & Suitor, C.W. (1998). Dietary reference intakes: the new basis 126. for recommendations for calcium and related nutrients, B vitamins, and choline. Journal of the American Dietetic Association, 98(6), 699-706. https://doi.org/10.1016/S0002-8223(98)00160-6
- Yi, J., Kebede, B.T., Dang, D.N.H., Buvé, C., Grauwet, T., Van Loey, A., Hu, X., & 127. Hendrickx, M. (2017). Quality change during high pressure processing and thermal processing of cloudy apple juice. Lwt, 75, 85-92. https://doi.org/10.1016/j.lwt.2016.08.041
- 128. Yin, X., Chen, K., Cheng, H., Chen, X., Feng, S., Song, Y., & Liang, L. (2022). Chemical stability of ascorbic acid integrated into commercial products: A review on bioactivity and delivery technology. Antioxidants, 11(1), 153. https://doi.org/10.3390/antiox11010153
- Yoruk, R., & Marshall, M.R. (2003). Physicochemical properties and function of plant polyphenol oxidase: a review 1. Journal of Food Biochemistry, 27(5), 361-422. https://doi.org/10.1111/j.1745-4514.2003.tb00289.x
- Younus, S.A., Mahdi, N.M., Al-Abbasy, O.Y., & Sheej Ahmad, O. (2025). Antioxidative effect of Maillard reaction products of spermine-sugar system on partially purified plum polyphenol Science and Technology (Iran), Journal of Food 22(160), https://doi.org/10.22034/fsct.22.160.227
- Yu, K., Zhou, L., Sun, Y., Zeng, Z., Chen, H., Liu, J., Zou, L., & Liu, W. (2021). Antibrowning effect of *Rosa roxburghii* on apple juice and identification of polyphenol oxidase inhibitors. Food Chemistry, 359, 129855. https://doi.org/10.1016/j.foodchem.2021.129855
- Zhang, Y., He, S., & Simpson, B. K. (2018). Enzymes in food bioprocessing—novel food enzymes, applications, and related techniques. Current Opinion in Food Science, 19, 30-35. https://doi.org/10.1016/j.cofs.2017.12.007
- Zhou, F., Xu, D., Liu, C., Chen, C., Tian, M., & Jiang, A. (2021). Ascorbic acid treatment 133. inhibits wound healing of fresh-cut potato strips by controlling phenylpropanoid metabolism. **Biology** Technology, 111644. **Postharvest** and 181, https://doi.org/10.1016/j.postharvbio.2021.111644
- 134. Zhu, J., Wu, H., & Sun, Q. (2019). Preparation of crosslinked active bilayer film based on chitosan and alginate for regulating ascorbate-glutathione cycle of postharvest cherry tomato (Lycopersicon esculentum). International Journal of Biological Macromolecules, 130, 584-594. https://doi.org/10.1016/j.ijbiomac.2019.03.006

مقاله مروري جلد ۲۱، شماره ٦، بهمن - اسفند ١٤٠٤، ص.

اسید اسکوربیک: ضرورتها و کاربردها در مهار قهوهای شدن آنزیمی میوهها و سبزیجات- یک بررسی جامع

عمر العباسي  $^{0}$  - زهرا الصراف $^{0}$  - نور مهدى  $^{0}$  - عليا البوواري  $^{0}$ 

تاریخ دریافت: ۱۴۰۳/۰۹/۲۱ تاریخ پذیرش: ۱۴۰۴/۰۳/۲۸

قهوهای شدن آنزیمی بیشتر در میوهها و سبزیجات تازه اتفاق میافتد و در تعیین ماندگاری محصول بسیار مهم است. دستهای از آنزیمها که بهعنوان پلیفنول اکسیداز شناخته میشوند، مسئول این تغییر رنگ هستند. پلیفنول اکسیداز آنزیم اصلی است که اکسیداسیون ترکیبات فنلی را در حضور اکسیژن کاتالیز می کند و رنگدانه قهوهای تشکیل میدهد. بنابراین، روشهای مختلفی برای جلوگیری از این واکنشها مورد نیاز است. اسید اسکوربیک طبیعی از جمله مهمترین مواد شیمیایی در متوقف کردن این واکنش و جلوگیری از قهوهای شدن است. این ماده یک جایگزین غیرسمی و مؤثر برای مواد شیمیایی مصنوعی است. همچنین با داشتن خواص آنتی اکسیدانی قوی و مهار رادیکالهای آزاد مشخص می شود. این مقاله مروری، مطالعات انجام شده در مورد استفاده از اسید اسکوربیک بهعنوان یک عامل ضد قهوهای شدن در منابع غذایی مختلف را خلاصه می کند. این مقاله ممکن است بینشی در مورد پتانسیل توسعه استفاده از اسید اسکوربیک در عملیات قهوهای شدن مواد غذایی و کاهش ضررهای اقتصادی در بخش مواد غذایی ارائه دهد.

واژههای کلیدی: اسید اسکوربیک، سبزیجات، کنترل قهوهای شدن، قهوهای شدن آنزیمی، میوهها

۱- گروه شیمی، دانشکده آموزش علوم محض، دانشگاه موصل، موصل، عراق

<sup>(\*-</sup> نویسنده مسئول: Email: chem.omar1978@uomosul.edu.iq)

۲- گروه شیمی دارویی، دانشکده داروسازی، دانشگاه نینوا، موصل، عراق

٣- اداره كل آموزش و پرورش نينوا، نينوا، عراق