#### Research Article

# Element Accumulations in Liver and Kidney Tissues of Some Bony Fish Species in the Southwest Caspian Sea

Masoud Sattari<sup>1,2\*</sup>, Mehdi Bibak<sup>1</sup>, Shima Bakhshalizadeh<sup>2</sup>, Mohammad Forouhar Vajargah<sup>1</sup>

<sup>1</sup> Department of Fisheries, Faculty of Natural Resources, University of Guilan, Sowmeh Sara, Iran <sup>2</sup> Department of Marine Sciences, Caspian Sea Basin Research Center, University of Guilan, Rasht, Iran

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#### Abstract

The Caspian Sea is the largest inland body of water in the world and so has both common characteristics of seas and lakes with over 153 fish species which inhabit the sea and its basin. However, little is known about the trace element (TE) contaminations (TECs) in its tissues. In the present study, 122 specimens of three fish species including Rutilus caspius (Roach, n=71), Leuciscus aspius (Asp, n=20), and Tinca tinca (Tench, n=31) were collected from three different fisheries regions (i.e. Astara, Anzali and Kiashahr) of the southern part of the Caspian Sea from September 2017 to June 2018. Inductively coupled plasma optical emission spectrometry (ICP-OES) was employed to measure TE levels in different fish tissues. An attempt was made to assess possible influences of habitat on element accumulation in the liver and kidney of three fish species in the southwest of the Caspian Sea basin. Some elements including Ca, K, Mg, P, S, Sc, and Sr showed different concentrations in the liver and kidney. Also their levels were significantly different between freshwater resident (Tench) and marine (Roach) species (p < 0.05). The differences among TECs in the liver and kidney of Roach, Asp and Tench were reduced to three components using principal component analysis (PCA). Results indicated that 83.60% of the total variability is related to TEs such as Cu, Fe, Sr, Ca, S, Na, Mg, K, and Al. The impact of habitat variability on the element accumulation was confirmed through linear chart obtained for liver and kidney (as body filtering organs) of Roach and Asp as marine residents as well as Tench as a freshwater resident. This could illustrate the borderline created by these habitats.

Keywords: Rutilus caspius, Leuciscus aspius, Tinca tinca, Trace elements, Caspian Sea

#### Introduction

The trace element (TE) pollution in water resources has long been found to be a serious environmental concern (Pagano et al., 2017; Capillo et al., 2018; Chorehi et al., 2013). Aquatic organisms can accumulate TEs in their bodies via respiration, adsorption and ingestion (Zhou et al., 2001; Boran et al., 2000). TE contamination is a serious problem in the coastal regions, due to waste disposal of discharges from agriculture, industries and some urban sources (Aliko et al., 2018; Burgos-Aceves et al., 2018). TE accumulation elevated in marine ecosystems as a direct result of anthropogenic activities (Seco-Gesto et al., 2007). TEs were categorized as potentially toxic (cadmium, arsenic, lead, mercury, nickel, etc.), probably essential (vanadium, cobalt) and essential (copper, selenium, iron, manganese, zinc) (Munoz-Olivas et al., 2001). Fish is considered as a suitable indicator for long term monitoring of TE contaminations in different water resources (Fazio et al., 2014; Sattari et al., 2019). Therefore, numerous studies have been conducted on TE accumulation in different fish species (Türkmen et al., 2007).

The Caspian Sea is the world's largest inland body of water and thus has characteristics common to both seas and lakes. It is bordered by Russia (Dagestan, Kalmykia, and Astrakhan oblasts), the Republic of Azerbaijan, Iran (Guilan, Mazandaran and Golestan provinces), Turkmenistan, and Kazakhstan (Vajargah et al., 2014; Sattari et al., 2019). Hence, not only it doesn't contain fresh water, but also it is under intense pollutant threats from industrial and agricultural effluents as well as growing urbanization in the most riparian countries of the Caspian Sea (Karrari et al., 2012).

There are numerous reports regarding the heavy metal contamination in aquatic environments of Iran including: Pourang et al., (2005) on five sturgeon species in the Caspian Sea; Abtahi et al., (2005) on *Liza aurata* in the south Caspian Sea; Sadeghirad, (2007) on *Acipenser persicus* and

<sup>&</sup>lt;sup>\*</sup>Corresponding author's e-mail address: *msattari@guilan.ac.ir* 

Acipenser stellatus from the Caspian Sea; De Mora et al., (2004) on coastal sediments from the Caspian Sea; Amini Ranjbar and Sotudehnia, (2005) on Mugil auratus of the Caspian Sea; Askary Sary and Beheshti, (2012) on Liza abu from the Karoun and Karkheh rivers; Beheshti, (2011) on Liza abu collected from the Karkheh and Karoon rivers; Ebrahimzadeh et al., (2011) on Liza saliens collected from the Caspian Sea; Khanipour et al., (2018) on Silurus glanis collected from Anzali Wetland, the southwest Caspian Sea; and Alipour and Banagar, (2018) on fish obtained from Gorgan Bay, the southeast Caspian Sea. There are also some reports on TE accumulations in R. kutum (Shahryari et al., 2010; Eslami et al., 2014; Mirzajani et al., 2016; Sattari et al., 2019). Eslami et al., (2011) reported the existence of TEs in muscle and liver of Perca fluviatilis and Tinca tinca in Anzali Wetland; Bibak et al., (2018) worked on heavy metal levels in sediments of the northern part of the Persian Gulf. This study aimed to determine the levels of some target trace elements (TEs) in the livers and kidneys of three fish species which were collected from the geographically different coastal regions of the Caspian Sea.

## **Materials and Methods**

This study was conducted in three fisheries regions including Kiashahr ( $37^{\circ} 42' 20''$  N,  $49^{\circ} 94' 95''$  E), Astara ( $38^{\circ} 42' 25''$  N,  $48^{\circ} 86' 87''$  E), and Anzali ( $37^{\circ} 46' 39''$  N,  $49^{\circ} 47' 99''$  E) along the south western coasts of the Caspian Sea. 122 specimens were collected from September, 2017 to January, 2018 from three different fish species including *Rutilus caspius* (Roach, n=71), *Leuciscus aspius* (Asp, n=20), and *Tinca tinca* (Tench, n=31) with gill net. The specimens were transported to the Fish Biology Laboratory, University of Guilan, Sowmeh Sara, Iran by a styrofoam cooler box at 4°C. Fish were washed using distilled water, dissected and pieces of muscle were dried in the

oven (80°C for 18 h) (Vajargah et al., 2018b). Fish age was determined with scales during the process. To extract TEs, 0.5 g of each tissue was digested in 10 ml of 65% nitric acid in a microwave oven. Then, specimens were passed through the Whatman filter paper No. 40 and were diluted in distilled water to the required volume. An inductively coupled plasma–optical emission spectrometry (ICP-OES) (Zarazma Co. Tehran, Iran) was employed to measure trace element levels in the specimens. Instrumental detection limits for trace and major (Al, Ca, Fe, K, Mg, Mn, Na and Si) element measurements were equal to 0.02 mgkg<sup>-1</sup>, and 0.1 mg kg<sup>-1</sup>, respectively.

### **Statistical analyses**

After examining the normality of acquired data and homogeneity of variances in the fish tissues (liver and kidney) from different habitats, the variability of TE concentrations was investigated through one-way analysis of variances (ANOVA). For heterogeneous variables, the Kruskal-Wallis test was employed, otherwise, we used Man-Whitney U test (Zar, 1996).

Principle component analysis (PCA) was used to reduce the number of variables without losing much information (Ouinn and Keouch. 2002). Eigenvalues against the number of principal components and also the values of cumulative variances were provided to define the important principle components and elements. Discriminant function analysis (DFA) was employed to calculate the exact place of each fish species which were Ward's related correctly. method, as а complementary method for DFA, was employed to construct cluster dendrogram using Euclidean distance (average linkage clustering). All statistical analyses were performed using SPSS version 16.0 (significance level  $\alpha$ =0.05, SPSS Inc., Chicago, IL, USA).



Figure 1. Map of the study area along the south western coasts of the Caspian Sea

#### Results

In the present study, several specimens from three different fish species were dissected and their kidney and liver tissues were examined for the presence of 36 elements including: Silver (Ag), Aluminum (Al), Arsenic (As), Barium (Ba) Beryllium (Be), Bismuth (Bi), Calcium (Ca), Cadmium (Cd), Cesium (Ce), Cobalt (Co), Chromium (Cr), Copper (Cu), Iron (Fe), Potassium (K), Lanthanum (La), Lithium (Li), Magnesium (Mg), Manganese (Mn), Molybdenum (Mo), Sodium (Na), Nickel (Ni), Phosphorus (P), lead (Pb), Rubidium (Rb), Sulfur (S), Antimony (Sb), Scandium (Sc), Silicon (Si), Tin (Sn), Strontium (Sr), Thorium (Th), Titanium (Ti), Uranium (U), Vanadium (V), Tungsten (W), Yttrium (Y) and Zinc (Zn).

Some elements such as Ca, K, Mg, P, S, Sc and Sr exhibited different concentrations between liver and kidney (\*p < 0.05, Table 1). Their levels were also displayed significant differences between Tench, as a freshwater resident, and Roach, as a marine species (Table 2). The variability of TECs in different edible tissues of Roach, Asp and Tench was reduced to three components using PCA (PC1= 46.32%, PC2 =22.48% and PC3=14.79%) (Figures 2-3). It was found that 83.60% of total variation is related to TEs such as Cu, Fe, Sr, Ca, S, Na, Mg, K and Al (Table 3). The three-dimensional diagram illustrated the weight of each component in PCA is shown in Figure 3 (Figure 3). The first component was mainly influenced by Na, Mg, K and S (Table 3). Ca and Sr had special contributions in PC2, while the highest values in PC3 were obtained for Cu, Fe and Al (Table 3).

**Table 1.** Mean of concentration $\pm$ SD for different elements in the liver and kidney of the sea water fish and lagoon fish of the Iranian Caspian Sea (\*p < 0.05).

Elements	<b>Elemental concentration (ppm; Median ± SD)</b>			<i>p</i> value	
	T. tinca	R. caspius	L. aspius		
Al	$019\pm0.20$	$0.22\pm0.11$	$0.05\pm0.05$	0.46	
Ca	$9.06\pm0.38$	$25.84 \pm 9.73$	$97.30 \pm 73.30$	0.02	
Cr	$0.00 \pm 0.00$	$0.02\pm0.01$	$0.09\pm0.01$	0.12	
Cu	$0.07\pm0.05$	$0.17\pm0.12$	$0.00\pm0.00$	0.36	
Fe	$1.59\pm0.59$	$4.82 \pm 1.92$	$0.63\pm0.25$	0.17	
Κ	$38.33 \pm 12.97$	$114.62\pm8.01$	$115.30\pm67.60$	0.03	
Mg	$4.03\pm0.81$	$15.06\pm2.33$	$17.05\pm5.25$	0.02	
Na	$20.57\pm5.78$	$56.16\pm6.25$	$48.50 \pm 19.70$	0.58	
Р	$36.27 \pm 11.60$	$117.00 \pm 13.94$	$220.60\pm119.9$	0.02	
S	$29.73 \pm 10.18$	$99.18 \pm 8.56$	$92.70\pm52.00$	0.03	
Sb	$0.08\pm0.08$	$0.04\pm0.02$	$0.09\pm0.09$	0.59	
Si	$0.00 \pm 0.00$	$0.30\pm0.06$	$0.26\pm0.06$	0.02	
Sr	$0.05\pm0.02$	$0.26\pm0.12$	$0.72\pm0.63$	0.02	
Zn	$0.70\pm0.12$	$2.08\pm0.28$	$1.20\pm0.30$	0.08	

**Table 2.** Pair-wise comparisons (Mann–Whitney U-test) of the significant elements among the fish belongs to sea water and lagoon of the Iranian Caspian Sea.

Elements	<b>Tench-roach</b>		Tench-asp		Roach-asp				
	M-W	Ζ	р	M-W	Ζ	Р	M-W	Ζ	р
Ca	0.00	-2.24	< 0.05	0.00	-1.73	>0.05	1.00	-1.55	>0.05
k	0.00	-2.24	< 0.05	1.00	-1.16	>0.05	5.00	0.00	>0.05
Mg	3.00	-2.24	< 0.05	1.00	-1.73	>0.05	0.00	-0.78	>0.05
Р	0.00	-2.24	< 0.05	0.00	-1.73	>0.05	3.00	-0.78	>0.05
S	0.00	-2.24	< 0.05	0.00	1.00	>0.05	1.00	5.00	>0.05
Sr	0.00	-2.24	< 0.05	0.00	-1.73	>0.05	4.50	-0.20	>0.05
Si	0.00	-2.29	< 0.05	0.00	-1.94	>0.05	4.50	-0.20	>0.05

M-W and Z are grouping variable scores for Mann–Whitney U and Kruskal–Wallis tests (\*p <0.05).



**Figure 2.** Principal component analysis (PCA) of elemental concentrations of the fish liver and kidney between habitats in the coastal water of the Iranian Caspian Sea. Scatter plots demonstrate individual fish scores for PC1 vs. PC2, PC1 vs. PC3, and PC2 vs. PC3 which together explain 83.60% of the total variance, this graph obtained by SPSS version 16.0.



**Figure 3.** Characteristic load for PC1, PC2 and PC3 obtained by multi-elemental principal components analysis (PCA) for the elemental concentrations of the fish liver and kidney between habitats in the south Caspian Sea water, this graph obtained by SPSS version 16.0.

**Table 3.** Characteristic load for PC1, PC2 and PC3 obtained by principal component (PCA) analysis for elemental concentrations of the fish liver and kidney between habitats (sea water and lagoon) in the water of Iranian Caspian Sea

<b>Elemental variables</b>	*PC1	PC2	PC3
Al	.012	.418	.629
Ca	.087	.903	361
Cr	.576	.661	360
Cu	.544	.084	.666
Fe	.615	.039	.683

K	.906	337	179
Mg	.965	.044	138
Na	.933	233	041
Р	.777	152	473
S	.952	267	078
Sb	287	.622	.286
Si	.844	.437	.188
Sr	.174	.931	199
Zn	.718	046	.256

\* Principle Components

The matrix composed of element concentrations in liver and kidney tissues of Roach, Asp, and Tench was described with two discriminant components. These experiments successfully discriminate the two investigated habitats (Wilk's Lambda=0.001,  $X^2$ =27.63, df=14 and *p*<0.05, Figure 4). Cluster

analysis, as a complementary method, divided the fish into two sub-groups. No variation was found for element concentrations in Roach and Asp, while Tench was placed in a distinct subgroup (Figure 5).



**Figure 4.** Plot of discriminant functions 1 and 2 for the elemental concentrations of the fish liver and kidney between habitats in the coastal water of the Iranian Caspian Sea, this graph obtained by SPSS version 16.0.

Dendrogram using Ward Method



**Figure 5.** Dendrogram derived from cluster analysis of the elemental concentrations of the fish liver and kidney between habitats in the coastal water of the Iranian Caspian Sea, Cluster analysis, as a complementary method, divided the fish into two sub-groups. No variation was found for element concentrations in Roach and Asp, while Tench was placed in a distinct subgroup, this graph obtained by SPSS version 16.0.

# Discussion

In the present study, an attempt was made to assess possible influences of habitat on the elements accumulation in the liver and kidney tissues of some fish species in the southwest of the Caspian Sea basin. Linear charts of element accumulations in the body filtering organs of Roach, Asp, and Tench exhibited their enough variability based on the habitats and also illustrated the borderline created by these habitats. The consequences of different elements bioaccumulation on fish tissues depend on sex, maturation stage, size, tissue type, habitat and fish diet (Azevedo et al., 2009). Previous studies revealed that element bioaccumulation in various fish tissues take place at different levels; but, in short time periods, filtering organs such as liver exhibit higher levels of these elements (Afonso et al., 2017, Alamdar et al., 2017; Salgado-Ramírez et al., 2017). This is while, gills and gut are the first organs receiving these elements (Tiphaine et al., 2018). Therefore, liver could be considered as the main organ for element aggregation monitoring studies (Salgado-Ramírez et al., 2017)

Since metal elements find their way to the aquatic environment and deposit in sediments, the demersal and benthivorous fish species are more susceptible to element bioaccumulation than planktivorous fish (Trevizani et al., 2019).

Little is known about Cu accumulation in fish species. However, it has been found in the higher than normal levels in the food chain which is distinguishable from low-level elements. It seems that raising in Metallothionein levels could be considered as an exact indicator of Cu existence in the ambient environment (Marijić and Raspor, 2007). This is reduced upon the migration of the fish from saltwater to freshwater (Ohji et al., 2007). It is also true for Sr which is found in higher amounts in saltwater in comparison to freshwater. So that, Strontium levels are higher in fish tissues with long residence times in sea water, regardless of the fish diet.

Meanwhile, there is a positive correlation between Sr and Ca. So that, its concentration is raised by low temperature and high salinity (Walther and Thorrold, 2006), instead of the fish diet in Asp tissues. Overall, the present study provides some basic information about elements bioaccumulation in the fish filtering tissues from different ecosystems with various salinity levels. These data could be applicable in determining the focal points of contaminations.

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# References

Abtahi B. and Sabbagh kashani A. (2005) Assessing Pb, Ni and Zn accumulation in the tissues of Liza aurata in the south Caspian Sea. Iranian Scientific Fisheries Journal 14: 65-78.

Afonso A., Gutiérrez A. J., Lozano G., González-Weller D., Rubio C., Caballero J. M., Hardisson A. and Revert C. (2017) Determination of toxic metals, trace and essentials, and macronutrients in Sarpa salpa and Chelon labrosus: risk assessment for the consumers. Environmental Science and Pollution Research 24(11): 10557-10569.

Alamdar A., Eqani S. A. M. A. S., Hanif N. Ali S. M., Fasola M., Bokhari H., Katsoyiannis I. A. and Shen H. (2017) Human exposure to trace metals and arsenic via consumption of fish from river Chenab, Pakistan and associated health risks. Chemosphere 168: 1004-1012.

Aliko V., Qirjo M. and Sula E. (2018) Antioxidant defense system, immune response and erythron profile modulation in Gold fish, Carassius auratus, after acute manganese treatment" Fish Shellfish Immunology 76:101–109.

Alipour H. and Banagar G. h. R. (2018) Health risk assessment of selected heavy metals in some edible fishes from Gorgan Bay, Iran. Iranian Journal of Fisheries Sciences 17(1): 21-34.

Amini Ranjbar G. h. and Sotudehnia F. (2005) Investigation of heavy metals accumulation in muscle tissue of Mugil auratus in relation to standard length, weight, age and sex. Iranian Scientific Fisheries Journal 14: 1-18. (In Persian).

Askary Sary A. and Beheshti M. (2012) Cadmium, iron, lead and mercury bioaccumulation in Abu Mullet, Liza abu, different tissues from Karun and Karkheh Rivers, Khozestan, Iran. Bulletin of Environmental Contamination and Toxicology 88: 158-161.

Azevedo J., Fernandez W., Farias L., Fávaro D. and Braga E. (2009) Use of Cathorops spixii as bioindicator of pollution of trace metals in the Santos Bay, Brazil. Ecotoxicology 18(5): 577-586. Beheshti M. (2011) Comparative study of concentration of heavy metals (Cu, Fe, Zn, Mn) in muscle, liver and gill organ of fish (Liza abu) in the Karoon and Karkheh rivers in Khuzestan province." M.Sc. Thesis. Islamic Azad University, Science and Research, Ahwaz. (In Persian).

Bibak M., Sattari M., Agharokh A., Tahmasebi S. and Imanpour Namin J. (2018) Assessing some heavy metals pollutions in sediments of the northern Persian Gulf (Bushehr province). Environmental Health Engineering and Management Journal 2018: 5(3): 175–179.

Boran M., Karacam H., Celikkale M. S. and Kose S. (2000) Levels of heavy metals in blue whiting caught from the eastern Black Sea area of Turkey. Toxicological and Environmental Chemistry 2000: 75, 67-73.

Burgos-Aceves M. A., Cohen A., and Smith Y. (2018) MicroRNAs and their role on fish oxidative stress during xenobiotic environmental exposures. Ecotoxicology and Environmental Safety. 148: 995-1000.

Capillo G., Silvestro S. and Sanfilippo M. (2018) Assessment of electrolytes and metals profile of the Faro Lake (Capo Peloro Lagoon, Sicily, Italy) and its impact on *Mytilus galloprovincialis*. Chemestry & Biodiversity.

Chorehi M. M., Ghaffari H., Hossaini S. A., Niazie E. H. N., Vajargah M. F. and Hedayati A. (2013) Acute toxicity of Diazinon to the Caspian vimba, *Vimba vimba persa* (Cypriniformes: Cyprinidae). International Journal of Aquatic Biology 1(6): 254-257.

De Mora S., Sheikholeslami M. R., Wyse E. and Azemard S. (2004) An assessment of metal contamination in coastal sediments of the Caspian Sea. Marine Pollution Bulletin 48: 61-77.

Ebrahimzadeh M. A., Eslami S. and Nabavi S. F. (2011) Determination of trace element level in different tissues of the leaping mullet (Lizasaliens, Mugilidae) collected from Caspian Sea. Biological Trace Element Research 144: 804-811.

Eslami S., Moghaddam A. H. and Jafari N. (2011) Trace element level in different tissues of *Rutilus frisii kutum* collected from Tajan River, Iran. Biological Trace Element Research 143: 965-973.

Eslami V., Sattari M. and Namin J.I. (2014) Concentration of heavy metals (Pb, Cd) in muscle and liver of Perca fluviatilis and *Tinca tinca* in Anzali Wetland, southwest the Caspian Sea. International Journal of Aquatic Biology 2(6): 319-324.

Fazio F., Piccione G. and Tribulato K. (2014) Bioaccumulation of heavy metals in blood and tissue of striped mullet in two Italian lakes. Journal of Aquatic Animal Health 26: 278–284.

Khanipour A. A., Ahmadi M. and Seifzadeh M. (2018) Study on bioaccumulation of heavy metals (cadmium, nickel, zinc and lead) in the muscle of wels catfish (*Silurus glanis*) in the Anzali Wetland. Iranian Journal of Fisheries Sciences 17(1): 244-250.

Marijić V. F. and Raspor B. (2007) Metallothionein in intestine of red mullet, *Mullus barbatus* as a biomarker of copper exposure in the coastal marine areas. Marine pollution bulletin 54(7): 935-940.

Munoz-Olivas R. and Camara C. (2001) Speciation related to human health, In: L, Ebdon, L, Pitts, R, Cornelis, H, Crews, OFX, Donad, P, Quevauviller, (Eds.), Trace Element Speciation for Environment, Food and Health. The Royal Society of Chemistry 331-353.

Ohji M., Arai T. and Miyazaki N. (2007) Comparison of organotin accumulation in the masu salmon *Oncorhynchus masou* accompanying migratory histories. Estuarine, Coastal and Shelf Science 72(4): 721-731.

Ohji M., Harino H. and Arai T. (2006) Differences in organotin accumulation among ecological migratory types of the Japanese eel Anguilla japonica. Estuarine, Coastal and Shelf Science 69(1-2): 270-290.

Pagano M., Porcino C. and Briglia M. (2017) The influence of exposure of cadmium chloride and zinc chloride on haemolymph and digestive gland cells from *Mytilus galloprovincialis*. International Journal of Environmental Research 11(2): 207-216.

Pourang N. and Dennis J. H. (2005) Distribution of trace elements in tissues of two shrimp species from the PersianGulf and roles of metallothionein in their redistribution. Environment International 31(3):325–341.

Sadeghirad M. (2007) Accumulation of trace elements in Acipenser persicus tissues in relation to feeding habits and mode of absorption of these pollutants. Iranian fisheries research organization. Internatinoal sturgeon research institut. 42P. Salgado-Ramírez C. A., Mansilla-Rivera I. and Rodríguez-Sierra C. J. (2017) Comparison of trace metals in different fish tissues of Scomberomorus spp.("sierra") and *Lutjanus synagris* ("arrayado") from Jobos Bay and La Parguera coastal areas in southern Puerto Rico. Regional studies in marine science 13: 1-11.

Sattari M., Imanpour J., Bibak M., Forouhar Vajargah M. and Khosravi A. (2019) Investigation of metal element concentrations in tissue of Rutilus frisii in the Southwest Caspian Sea. Iranian Scientific Fisheries Journal 28(3): pp.149-161.

Sattari M., Namin J. I., Bibak M., Vajargah M.F., Bakhshalizadeh S. and Faggio, C. (2019) Determination of Trace Element Accumulation in Gonads of Rutilus kutum (Kamensky, 1901) from the South Caspian Sea Trace Element Contaminations in Gonads. Proceedings of the National Academy of Sciences, India Section B: Biological Sciences

Sattari M., Namin J. I., Bibak M., Vajargah M. F., Faggio C. and Haddad M. S. (2019) Trace and macro elements bioaccumulation in the muscle and liver tissues of *alburnus chalcoides* from the south Caspian Sea and potential human health risk assessment. Journal of Energy, Environmental & Chemical Engineering 4(1):13.

Sattari M., Namin J. I., Bibak M., Vajargah M.F., Hedayati A., Khosravi A. and Mazareiy M.H. (2019) Morphological comparison of western and eastern populations of Caspian kutum, Rutilus kutum (Kamensky, 1901)(Cyprinidae) in the southern Caspian Sea. International Journal of Aquatic Biology 6(4): 242-247.

Seco-Gesto E. M., Moreda-Piñeiro A. and BermejoBarrera A. (2007) Multi-element determination in raft mussels by fast microwave assisted acid leaching and inductively coupled plasma-optical emission spectrometry, Talanta. 72: 1178.

Shahryari A., Golfirozy K., Noshin S. (2010) Muscular concentration of cadmium and lead in carp, mullet and kutum of the Gorgan Bay, Caspian Sea. Iranian Journal of Fisheries Sciences 19(2): 95-100.

Tiphaine M., Pierre C., Tiphaine C., Paco B., Christophe B.P., Sandrine B., Emmanuelle R. and Marc B. (2018) Trace metal concentrations in the muscle of seven marine species: Comparison between the Gulf of Lions (North-West Mediterranean Sea) and the Bay of Biscay (North-East Atlantic Ocean). Marine pollution bulletin 135: 9-16.

Trevizani T. H., Domit C., Vedolin M. C., Angeli J. L. F. and Figueira R. C. L. (2019) Assessment of metal contamination in fish from estuaries of southern and southeastern Brazil. Environmental monitoring and assessment 191(5): 308.

Türkmen M., Türkmen A. and Tepe Y. (2007) Metal contaminations in five fish species from Black, Marmara, Aegean and Mediterranean seas, Turkey. Journal of the Chilean Chemical Society 52: 4, 1314-1318.

Tzeng W., Shiao J., and Iizuka Y. (2002) Use of otolith Sr: Ca ratios to study the riverine migratory behaviors of Japanese eel Anguilla japonica. Marine Ecology Progress Series 245: 213-221.

Vajargah M. F., Hedayati A., Yalsuyi A. M., Abarghoei S., Gerami M. H. and Farsani, H. G. (2014) Acute toxicity of butachlor to caspian kutum (*Rutilus frisii Kutum* Kamensky, 1991). Journal of Environmental Treatment Techniques 2(4): 155-157.

Walther B. D. and Thorrold S. R. (2006) Water, not food, contributes the majority of strontium and barium deposited in the otoliths of a marine fish. Marine Ecology Progress Series 311: 125-130.

Zhou J. L., Salavador S. M. and Liu Y. P. (2001) Heavy metals in the tissues of common dolphins (*Delphinus delphis*) stranded on the Portuguese coast. Science of the Total Environment 273: 61.

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