

# *In silico* Identification of a Putative lncRNA-miRNA Regulatory Network Associated with Mucin Regulation in Colon Cancer

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## Abstract

Colorectal cancer is one of the most common cancers and is one of the leading causes of cancer-related deaths worldwide. The underlying biological mechanisms for the development of colorectal cancer are largely unidentified. Several genes likely involved in the pathogenesis of colorectal cancer have been identified. However, some other genes might have less evident functions. One gene family with prominent functional roles in the normal colon is mucin. Multiple studies have demonstrated the involvement of mucins in the pathogenesis of human malignancies. Therefore, due to the lack of an inclusive investigation of mucins' expression, mechanism of action, and involvement in colon adenocarcinoma's underlying biology, diagnosis, and prognosis, we sought to unearth their potential involvement and related regulatory networks in this disease. In this investigation, a step-wise manner was used, and a plethora of databases and algorithmic tools were applied. Due to a significant upregulation at both mRNA and protein levels and following a thorough evaluation of diagnostic and prognostic values in colon adenocarcinoma, MUC13 was determined to be the most relevant regulatory mucin in colon carcinoma. Altogether, these findings indicate a putative ncRNA-mRNA network, including hsa-mir-136-5p, hsa-mir-27a-3p, NEAT1, and XIST, to be involved in regulating MUC13 in colon cancer. This step-wise investigation implies that MUC13 may have a crucial role in the underlying molecular mechanisms for the initiation or progression of colon cancer. In addition, it provides insights into molecular mechanisms and possible regulatory non-coding RNA networks that might be responsible for regulating MUC13 expression.

**Keywords:** colon cancer, lncRNAs, miRNAs, mucins, MUC13, regulatory networks

## Introduction

Globally, over 1.9 million new cases of colorectal cancer were diagnosed in 2020, resulting in approximately 935,000 deaths. This cancer accounts for 10% of all cancer cases and cancer-related mortalities. Colorectal cancer is the second most common cause of cancer-related mortality and the third most common cancer type worldwide (Sung et al., 2021). Considering the high incidence and deaths caused by this cancer, many of its risk factors, mainly genetic predisposition characteristics, have not been identified. Thus, this

malignancy's molecular mechanisms need further investigation (Dekker et al., 2019).

Numerous studies have revealed the involvement of abnormal genes and proteins that play important roles in the initiation and progression of colorectal cancer. Mucins are a family of high molecular weight and heavily glycosylated proteins that are extensively expressed in mucosal tissues. Despite common structural characteristics, such as densely O-glycosylated filamentous domains, several types of mucins exist with distinct activities. Mucins have significant functions in multiple biological processes, including homeostasis, signaling, and cell protection (Corfield, 2015). Some mucins' aberrant expression and abnormal structure and function

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contribute to the underlying properties related to tumorigenesis and progression. However, their roles in the development of colon cancer are poorly characterized (Wi et al., 2021). Interestingly, there is a subtype of colorectal cancer that is characterized by the abundant expression of extracellular mucin known as mucinous colorectal cancer (Luo et al. 2019a).

Non-coding genes comprise the majority of the human genome. Numerous reports indicate that non-coding RNAs (ncRNAs) play pivotal roles in the biology of normal and cancerous cells (Yan & Bu, 2021). There are several types of ncRNAs, of which long non-coding RNAs (lncRNAs) and microRNAs (miRNAs) have received much attention. By definition, lncRNAs are non-coding RNAs with over 200 nucleotides that lack protein-coding ability (Yarmishyn & Kurochkin, 2015). Under physiological circumstances, lncRNAs play a major role in diverse biological processes such as transcription, splicing, epigenetic regulation of gene expression, and modification of chromatin structure. Dysregulation of lncRNA expression has been reported to contribute to the initiation, progression, and metastasis of various types of cancer, including colon, hepatocellular, and breast cancers (Marchese et al., 2017). On the other hand, microRNAs are short non-coding RNAs consisting of ~22 nucleotides that serve crucial roles in various biological pathways. In normal physiological circumstances, miRNAs operate through feedback mechanisms to protect critical biological processes, such as cell apoptosis, differentiation, and proliferation. Research findings demonstrate that miRNA expression is abnormal in human malignancies (Reddy, 2015) and the interplay between lncRNAs and miRNAs plays a crucial role in controlling the expression and functionality of proteins in cancer. The competing endogenous RNA (ceRNA) theory presented by Salmena and his colleagues states that some lncRNAs with a spongy-like action counteract the inhibitory effects of miRNAs on gene expression (Salmena et al., 2011). Based on this theory, some lncRNAs by sequestering specific microRNAs and RNA-binding proteins lead to altered levels of target mRNAs. There is substantial research showing that the ceRNA networks are involved in a broad range of human malignancies, such as colon (Sun et al., 2020), liver (Wang et al., 2017), gastric (X. Z. Yang et al., 2018), and lung (Wang et al., 2020) cancers.

Previous studies have demonstrated that some lncRNAs, such as *XIST* and *NEAT1*, are aberrantly expressed in colorectal cancer. This aberrant expression is associated with tumorigenesis, tumor

progression, metastasis, and an unfavorable prognosis. Due to their involvement in colon cancer progression, lncRNAs may act as valuable biomarkers for detecting or predicting the disease (Liu et al., 2020; Sun et al., 2018). Nevertheless, the present knowledge on the most influential lncRNA-miRNA-mRNA networks that are highly associated with colon cancer prognosis is ambiguous.

In this study, we examined the differential expression of mucins in colon cancer tissue compared to healthy tissues. We selected candidate mucins with the highest possibility of having functional roles in colon cancer and subsequently predicted miRNAs and lncRNAs that are associated with these mucins. Based on these findings and the expression of related lncRNAs and miRNAs, we identified lncRNAs and miRNAs that could be used to construct regulatory networks related to these candidate mucins. In addition, we assessed their potential for prognostic and diagnostic purposes. Ultimately, we constructed novel regulatory networks that seem to be significantly associated with colon cancer. This study sheds light on the potential roles of non-coding RNAs in regulating mucins in colon cancer. These RNAs might have the value of being potential therapeutic targets or diagnostic biomarkers. Further analysis and validation of these findings in a laboratory setting may contribute to a better understanding of the development and progression of colon cancer.

## Materials and Methods

The high-throughput mass spectrometry data for colon cancer was acquired from the Clinical Proteomic Tumor Analysis Consortium (CPTAC) incorporated in the UALCAN web portal (<http://ualcan.path.uab.edu/index.html>). UALCAN is a web portal to perform in-depth analysis of gene expression data in cancer. This data consisted of 97 samples of colon cancer, including different stages of cancer ranging from stages I through IV, and 100 normal control samples. Each sample's downloaded protein expression values were log<sub>2</sub> normalized, and Z-values were calculated (Chandrashekar et al. 2017). Then, using the GEPIA2 web server, we retrieved the values for the quantity of mucin gene family transcripts in colon adenocarcinoma. The GEPIA2 (Gene Expression Profiling Interactive Analysis 2) web server is a database that represents the gene expression analysis using tumor and normal data from TCGA and the GTEx projects, respectively (Tang et al. 2019). The analysis of gene expression in colon adenocarcinoma embedded in this database was conducted using 275 tumor samples (stages I to IV) and 349 normal controls. We

identified differentially expressed genes in colon adenocarcinoma, taking  $\log_2FC > 1$  as the threshold criterion for the analysis. In addition, we considered a false discovery rate adjusted  $p$ -value  $< 0.05$  as the statistically significant cutoff. We used the LIMMA (linear models for microarray data) package which utilizes the linear model and Bayesian empirical method for analysis as the differential analysis method (Ritchie et al., 2015).

Afterward, the diagnostic value of *MUC13* was evaluated using the ROC curve with the TCGA colon cancer and normal colon expression data, and statistical significance was defined as an adj.  $p$  value  $< 0.05$ . Then, six target gene prediction tools, including PicTar, miRDB, MicroCosm, miRanda, PITA, and TargetScan, which are algorithms for identifying microRNA targets, were utilized to predict miRNAs that potentially interact with *MUC13*. Four principal elements of the miRNA-mRNA target interaction are common characteristics on which target prediction tools are based: site accessibility, free energy, seed match, and conservation. All the predicted miRNAs were used in the subsequent filtering steps. We identified differentially expressed miRNAs (DEMs) in colon adenocarcinoma using dbDEMC (Xu et al. 2022). The experiment for this purpose included 444 cases and 8 controls, respectively. The dbDEMC is designed to provide a systematic resource for storing and querying DEMs in human cancers. The expression values were subjected to logarithmic transformation and quantile normalization. Then, the LIMMA package was used to select miRNAs with a significant difference between their mean expression level in case and control samples, with a false discovery rate (FDR) of less than 0.05. We extracted miRNA-seq expression data from case-control design experiments.

After that, for the prediction of lncRNA-*MUC13* interactions, we used the RIBlast system, a high-speed method based on the seed-and-extension algorithm (Fukunaga and Hamada 2017). RIBlast identifies seed regions by employing suffix arrays and then applies an RNA secondary structure energy model to extend the seed regions. The prediction of local base pair interactions is based on interaction energies calculated from accessibility and hybridization energies. We used a -16 kcal/mol score as the interaction energy threshold.

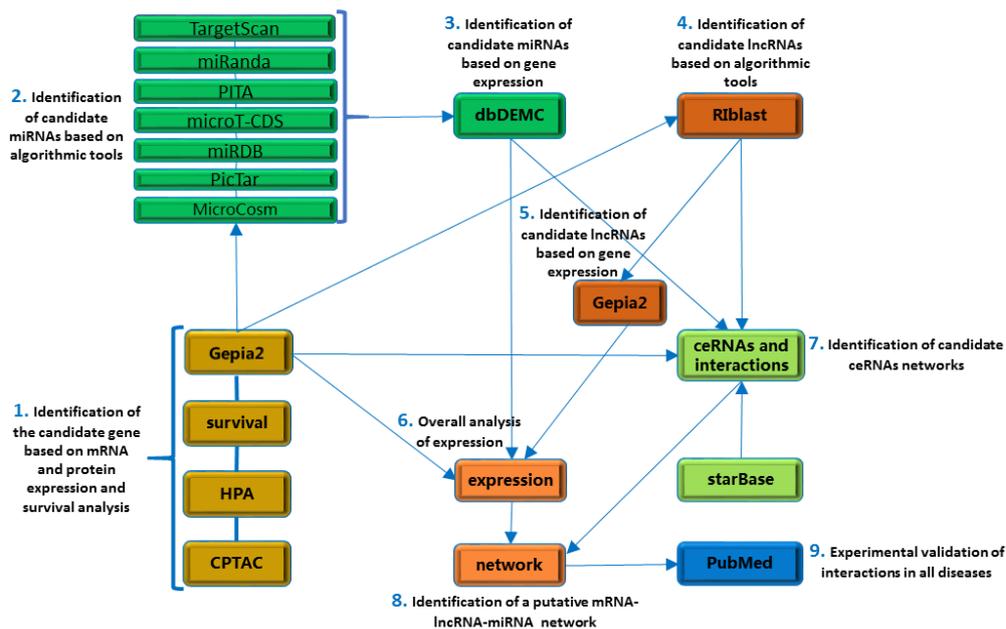
Then, we utilized starBase v2.0, a platform for studying RNA interactions, to decode miRNA-lncRNA interaction networks from CLIP-Seq data (Li et al. 2014). In the final step, to produce a more reliable prediction of interactions for constructing regulatory networks, we conducted a manual search of the PubMed database to extract data from validation experiments. PubMed is a public database that offers access to numerous abstracts and references related to biomedicine and the life sciences. In this regard, we queried this database for related lncRNA-miRNA interactions.

## Results

The present research involves multiple sequential stages, including aggregation and exploration of RNA, miRNA, and protein expression data, curve analyses for survival and receiver operating characteristics, and multiple *in silico* analyses. The main objective of this study was to detect potential non-coding networks associated with members of the mucin gene family in colon cancer. A schematic diagram of the study for *MUC13* is represented in Figure 1.

### Differential Expression Analysis of Mucins in Colon Cancer

In the first step, the protein expression of mucins in colon cancer was assessed using CPTAC data. The expression of *MUC13* protein in colon cancer samples was remarkably upregulated, whereas *MUC2*, *MUC5B*, and *MUC4* expressions were downregulated compared to normal samples (Figure 2A). The expression level of *MUC13* protein progressively increased from the early to advanced stages of cancer (Figure 2B). Immunohistochemical staining of *MUC13* protein confirmed higher expression in colon cancer tumors than in normal colon tissues (Figure 2C). On the other hand, the protein expression of *MUC12*, *MUC6*, *MUC5AC*, and *MUC1* in colon cancer samples compared to that in normal samples was not significantly changed (data not shown). Unfortunately, no protein expression information was available for the rest of the mucin family members in colon cancer.



**Figure 1.** The workflow of the study for predicting and identifying novel non-coding networks related to *MUC13* regulation in colon cancer. In this regard, databases including Gepia2, CPTAC, HPA, dbDEMC, starBase, and PubMed were used, and algorithmic tools, including TargetScan, miRanda, PITA, microT-CDS, miRDB, PicTar, MicroCosm, and Riblast were applied.

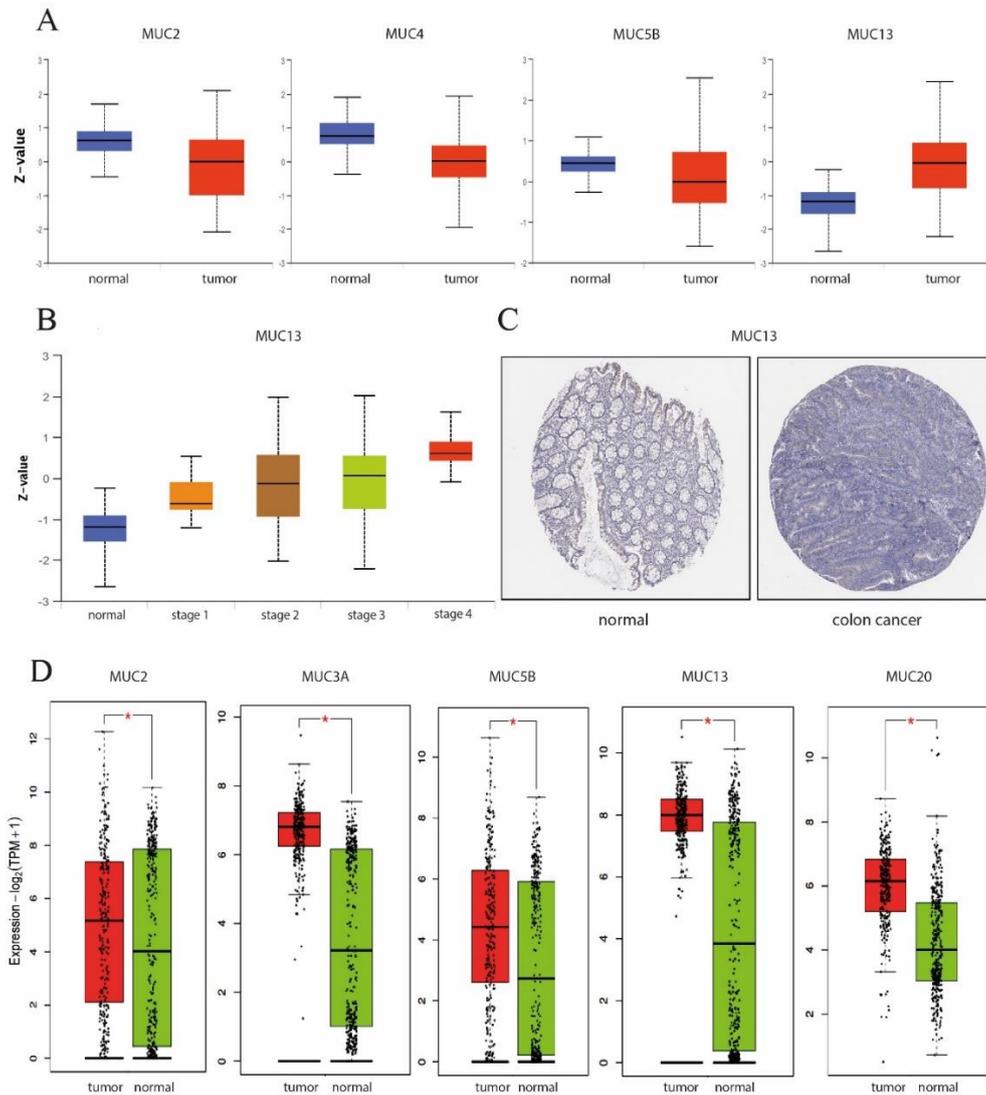
This expression data was first  $\log_2$  (Transcripts Per Million (TPM)+1) transformed for differential analysis, and the  $\log_2$ FC was defined as the median. Genes with higher  $\log_2$ FC values and lower  $q$  values than pre-set thresholds were considered differentially expressed genes. *MUC13*, *MUC2*, *MUC3A*, *MUC5B*, and *MUC20* genes were found to have significantly higher expressions in colon adenocarcinoma compared to normal pairs (Figure 2D). These mucin family members identified as differentially expressed are shown in Table 1. Consistent with the findings obtained from the CPTAC and considering the significant differential expressions, *MUC13* was selected as the most significant member of the mucin gene family in colon cancer for further analysis.

### The Prognosis and Diagnosis Significance of Candidate Mucins in Colon Cancer

We aimed to determine whether the expression changes of *MUC2*, *MUC3A*, *MUC5B*, *MUC13*, and *MUC20* in colon cancer could have promising predictive values for the prognosis of this disease. Kaplan-Meier plots were used to evaluate the prognostic significance of *MUC2*, *MUC3A*, *MUC5B*, *MUC13*, and *MUC20* in colon cancer. The Kaplan-Meier curves were retrieved from Gepia2,

and analysis was performed based on the expression status of mucins. In this section, we chose two indices: relapse-free survival and overall survival, and log-rank and hazard ratio tests were utilized for hypothesis tests. The overall survival and relapse-free survival analyses based on colon cancer did not show a significant prognostic impact for the increased expression of candidate mucins. However, in the case of *MUC13*, the difference between the two groups' median survival time was considerable (Figure 3B).

Based on the expression status of *MUC13* in colon cancer, we investigated whether it could serve as a reliable predictor for the diagnosis and prognosis of this malignancy. We assessed the diagnostic significance of this mucin in colon cancer using receiver operating characteristic curve analysis. The result of the ROC curve analysis showed that *MUC13* can be used to differentiate between colon cancer cases and normal controls (Figure 3A). Considering all evaluated features, *MUC13* appears to be the most promising member of the mucin gene family in colon cancer.



**Figure 2.** Screening the expression status of mucins in colon cancer. A. The Differential expression of mucin proteins in colon cancer, retrieved from CPTAC data. B. Proteomic expression profile of *MUC13* based on the individual cancer stages. C. The expression of *MUC13* in colon cancer and normal tissue samples extracted from HPA; antibody: HPA079663. D. Differentially expressed mucin gene family members in colon cancer, extracted from Gepia2. Only statistically significant results are shown.\* $p < 0.05$

**Table 1.** Mucin family members with significantly high expression in colon adenocarcinoma

Gene symbol	Median (Tumor)	Median (Normal)	logFC	adj. $p$ -value
<i>MUC5B</i>	20.540	5.640	1.698	9.61e-8
<i>MUC3A</i>	110.990	8.300	3.590	1.14e-64
<i>MUC13</i>	255.043	13.420	4.150	4.83e-53
<i>MUC20</i>	69.589	15.070	2.135	6.01e-39
<i>MUC2</i>	35.009	15.230	1.150	3.08e-2

**Prediction and Analysis of MUC13-Non-coding Interactions in Colon Cancer**

We utilized several gene interaction prediction tools to identify miRNAs that probably interact with MUC13. These tools were microT-CDS, miRDB, PicTar, MicroCosm, TargetScan, miRanda, and PITA. As a result, a total of 127 miRNAs were obtained based on algorithmic computations. A large number of these miRNAs were excluded based on their differential expressions. The expression profiles of these miRNAs in COAD were assessed.

**Table 2.** List of predicted miRNAs with significant changes in expression

Symbol	average expression	logFC	adj. p-value
hsa-miR-136-5p	4.278	5.818	2.33E-135
hsa-miR-139-5p	4.336	-5.238	4.23E-53
hsa-miR-7-1-3p	4.782	4.524	1.25E-34
hsa-miR-324-3p	3.620	-3.392	4.94E-28
hsa-miR-361-3p	6.257	-3.050	2.34E-23
hsa-miR-324-5p	3.942	-1.967	5.53E-13
hsa-miR-27a-3p	10.470	2.001	2.42E-11
hsa-miR-4723-3p	-2.393	-0.544	5.55E-12
hsa-miR-132-3p	6.139	-2.147	7.50E-11
hsa-miR-23a-5p	1.583	-2.967	1.38E-09
hsa-miR-668-3p	-2.220	-0.953	6.19E-09
hsa-miR-143-3p	16.513	2.528	1.24E-07
hsa-miR-6862-5p	-2.423	-0.337	2.60E-08
hsa-miR-1193	-2.414	-0.321	6.58E-08
hsa-miR-6764-5p	-2.144	-0.617	1.64E-05
hsa-miR-624-5p	0.073	0.576	0.000494914

For this purpose, we used dbDEMC. Seventeen of these miRNAs had significant differential expression (Table 2), and five of these miRNAs were significantly upregulated.

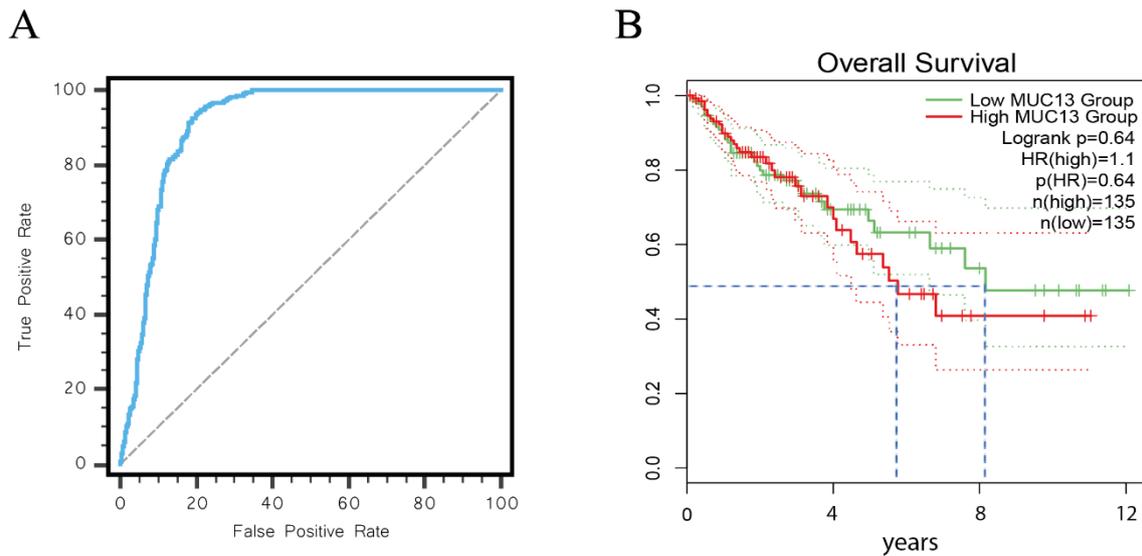
To investigate the molecular interactions of MUC13 in colon cancer, we used RIBlast to predict lncRNAs that may potentially interact with MUC13. After specifying MUC13 as a gene of interest, 92 lncRNAs were predicted by computational prediction. Subsequently, we evaluated the expression levels of the predicted lncRNAs using Gepia2, and seven lncRNAs with significant differential expression were identified (Table 3). The correlation between lncRNAs and MUC13 was assessed in colon cancer data.

**Table 3.** List of predicted lncRNAs with significant changes in expression

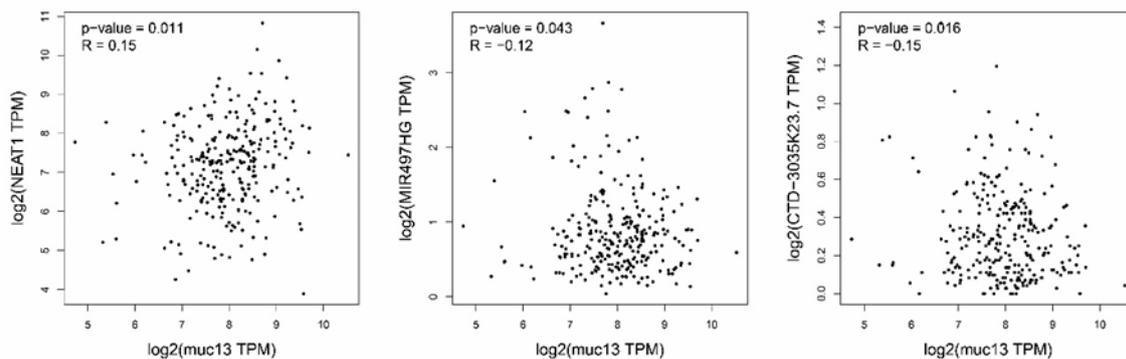
Symbol	Median (Tumor)	Median (Normal)	logFC	adj. p-value
NEAT1	153.915	1060.404	-2.776	5.92E-75
CTD-253719.12	5.1	18.581	-1.683	3.36E-48
CTD-3035K23.7	0.19	1.970	-1.319	4.31E-67
XIST	0.21	3.890	-2.015	9.66E-06
CTD-3193O13.11	1.39	0.160	1.043	6.01E-65
MIR497HG	0.69	9.910	-2.691	1.55E-124

**Construction of the Conceptual Regulatory Network**

To identify interactions with the highest chance of occurring in our pool of genes and to acquire more reliable interactions based on validated experiments, we obtained validated interactions between our candidate lncRNAs and microRNAs by querying PubMed and starBase. We identified only two microRNAs and two lncRNAs associated with our candidate gene, MUC13. Finally, we put together interactions between candidate miRNAs and lncRNAs and constructed potential lncRNA-



**Figure 3.** The prognostic and diagnostic values of *MUC13* in colon cancer. (A) The ROC curve of *MUC13* in colon cancer, AUC = 0.909,  $p < 0.001$ . (B) The overall survival curve of *MUC13* in colon cancer.



**Figure 4.** Scatterplots displaying the correlation of *MUC13* with different lncRNAs.  $p$ -value  $< 0.05$ .

miRNA-mRNA axes (including two lncRNAs, *NEAT1* and *XIST*, and two miRNAs, hsa-mir-136-5p and hsa-mir-27a-3p) that may be involved in the regulation of *MUC13* in colon cancer.

## Discussion

Mucins are involved in tumor initiation, progression, and metastasis. Aberrant localization and over-expression of *MUC13* in colon cancer are known. Based on the step-wise investigation, we determined *MUC13* as the candidate mucin. Then, we recognized candidate miRNAs (hsa-mir-136-5p and hsa-mir-27a-3p) and lncRNAs (*NEAT1* and *XIST*) related to *MUC13* in a reverse step-wise manner based on algorithmic prediction, expression, and validated interactions. Finally, we constructed

novel non-coding RNA networks consisting of these non-coding RNAs related to *MUC13*.

*MUC13*, which has two subunits, works as a transmembrane protein and is expressed in the normal colon. There are inconsistencies in the reports of *MUC13* expression in colon cancer. Some studies, such as those conducted by Gupta et al. and Walsh et al., indicated that *MUC13* expression in colon cancer is comparable to or higher than in normal samples (Gupta et al. 2014; Walsh et al. 2007). Sheng et al. found a significant association between high cytoplasmic *MUC13* expression and tumor grade in colorectal cancer tumors (Sheng et al. 2017). Likewise, Sheng et al. reported that higher levels of *MUC13* expression in colorectal tumors were associated with poor survival outcomes in their study patient cohorts (Sheng et al. 2019). In a newer study, Sojka et al. confirmed that higher levels of *Muc13* expression in tumor tissue in colorectal

cancer patients would have inferior survival compared to those with lower levels of expression (Sojka et al. 2023). Other studies, such as those carried out by Williams et al. and Packer et al., documented a decrease in *MUC13* expression in cases of colon cancer (Packer et al. 2004; Williams et al. 2001). Our findings align with the former group of studies. These variations could be attributed to dissimilarities in the sources of the samples or the types of control groups utilized. Gupta et al. studied the function of *MUC13* in colon cancer and found that overexpressed *MUC13* has promoting effects on tumorigenesis and metastasis through oncogenes and impacts on several pathways (Gupta et al. 2014). Sheng et al. proposed that *MUC13* has protective activities in colorectal cancer cells against cell death by activating the NF- $\kappa$ B pathway and suggested it as a gene of high importance in this malignancy (Sheng et al. 2017). Sheng et al. suggested that *MUC13* stimulates the expansion of colitis-associated colorectal tumors through  $\beta$ -catenin activity (Sheng et al. 2019).

We hypothesized that interactions among dysregulated non-coding RNAs, collectively constituting a regulatory network, impact the dysregulated activity and expression of *MUC13* in colon cancer. Thus, using several algorithms and databases, we constructed a regulatory network for *MUC13* in colon cancer. Candidate microRNAs were first identified using several algorithmic tools to investigate the potential non-coding regulatory network of *MUC13* systematically. Then, based on the ceRNA theory and combined with the differential expression status of miRNAs in colon adenocarcinoma and validated interactions, we selected two candidate miRNAs out of 17. After that, two lncRNAs were selected based on their expression and structural profiles. One of the candidate microRNAs in this study was hsa-mir-136-5p. Several studies have investigated its role in different types of cancer, including colorectal cancer. Jin et al. found that over-expressed hsa-miR-136-5p plays a significant role in controlling *SOX9*, thereby facilitating the progression of colorectal cancer (Jin et al. 2023).

The second candidate microRNA was hsa-mir-27a-3p. The oncogenic roles of hsa-mir-27a-3p in colon cancer have already been reported. Liang and colleagues found that hsa-miR-27a-3p expression was increased in CRC tissues and significantly correlated with survival, histological differentiation, and metastasis. Their study demonstrated that the hsa-miR-27a-3p/RXR $\alpha$ /Wnt/ $\beta$ -catenin pathway is implicated in the progression of CRC. MiR-27a-3p targets RXR $\alpha$ , which contributes to the activity of

the Wnt signaling pathway during the progression of CRC (Liang et al. 2017). Su et al. reported that, through the targeting of BTG1, hsa-miR-27a-3p regulates apoptosis and proliferation in colon cancer. They found that the impacts of hsa-miR-27a-3p on colon cancer cell apoptosis and proliferation were comparable to those of the BTG1, which acts as a tumor suppressor gene (Su et al. 2019). Another study by Chen et al. showed that lncRNA RMST inhibits colorectal cancer progression by inactivation of the Wnt signaling pathway and competitive interaction with the hsa-miR-27a-3p and RXR $\alpha$  (Chen et al. 2023).

lncRNAs have pivotal roles in many characteristics of cancer, such as apoptosis, migration, cell proliferation, and invasion (Huarte 2015). After a multi-step investigation, we determined *XIST* to be one of our candidate lncRNAs. There is increasing evidence that the lncRNA X-inactive specific transcript (*XIST*) plays a critical role in developing and regulating cell growth. Besides the primary function of *XIST* in dosage compensation of X chromosome, this lncRNA is also involved in the pathogenesis of cancer and many other human diseases by acting as a competing endogenous RNA (ceRNA) (Wang et al. 2021). According to a study by Yang et al., *XIST* functions as a competing endogenous RNA for miR-93-5p and promotes colorectal cancer progression via the HIF-1A/AXL signaling pathway (Yang et al. 2020).

The other lncRNA identified as a candidate in our investigation was *NEAT1*. The lncRNA *NEAT1* (Nuclear Enriched Abundant Transcript 1) performs a crucial role in the assembly and stability of paraspeckles and has been widely recognized as an essential element of paraspeckles. *NEAT1* promotes the development and progression of cancer by altering the levels of expression of genes that participate in several properties of cancer, such as growth, metastasis, invasion, regulation of tumor cells, and migration (Dong et al. 2018). Luo et al. reported that the lncRNA *NEAT1*, functioning as a competing endogenous RNA, contributes to colorectal cancer development by binding miR-34a competitively to SIRT1 and upregulating the Wnt/ $\beta$ -catenin signaling pathway (Luo et al. 2019b).

According to the competitive endogenous RNA hypothesis, some lncRNAs have regulatory roles on downstream protein-coding genes by sequestering miRNAs. For this reason, we performed further investigations to identify potential lncRNA-miRNA interactions between the candidate miRNAs and lncRNAs. Interestingly, several of the interactions we proposed in this network have been validated in

previous studies but not necessarily in colon cancer. Jiang and colleagues reported that the long non-coding RNA *NEAT1* elevates hypoxia-induced apoptosis of renal tubular epithelium by downregulating miR-27a-3p. Inhibition of *NEAT1* expression reduced miR-27a-3p overexpression, leading the authors to conclude that *NEAT1* negatively regulates the expression of hsa-miR-27a-3p. Dong et al. documented that *NEAT1* promotes Alzheimer's disease development by downregulating has-mir-27a-3p (Dong et al. 2021). The authors employed dual luciferase reporter gene and RNA pull-down assays to evaluate the potential interaction between the lncRNA hsa-mir-27a-3p and *NEAT1* and concluded that *NEAT1* targets hsa-mir-27a-3p. These researchers concluded that *NEAT1* contributes to the development of Alzheimer's disease via downregulating hsa-mir-27a-3p. Zhang and colleagues applied luciferase reporter and RNA immunoprecipitation assays to demonstrate that *XIST* can interact with hsa-mir-27a-3p. They suggested that long non-coding RNA *XIST* affects cerebral ischemia/reperfusion injury by regulating miR-27a-3p/FOXO3 signaling (Zhang et al. 2021).

In conclusion, non-coding regulatory interactions and networks are receiving increasing attention in cancer research, leading to a deeper understanding of cancer biology and subsequent applications. The constructed non-coding regulatory networks are proposed as novel networks with potential oncogenic roles in colon cancer. Here, we constructed potential regulatory networks of miRNAs and lncRNAs related to *MUC13* in colon

cancer utilizing a step-by-step reverse identification from mRNA to miRNA and lncRNA (Figure 5). These findings might lead to novel insights into the molecular biology of colon cancer. However, it should be noted that the conduct of experimental validation is imperative. Subsequent investigations should concentrate on elucidating the regulatory role of the predicted *MUC13*-related non-coding axes in colon cancer through in vitro and in vivo investigations.

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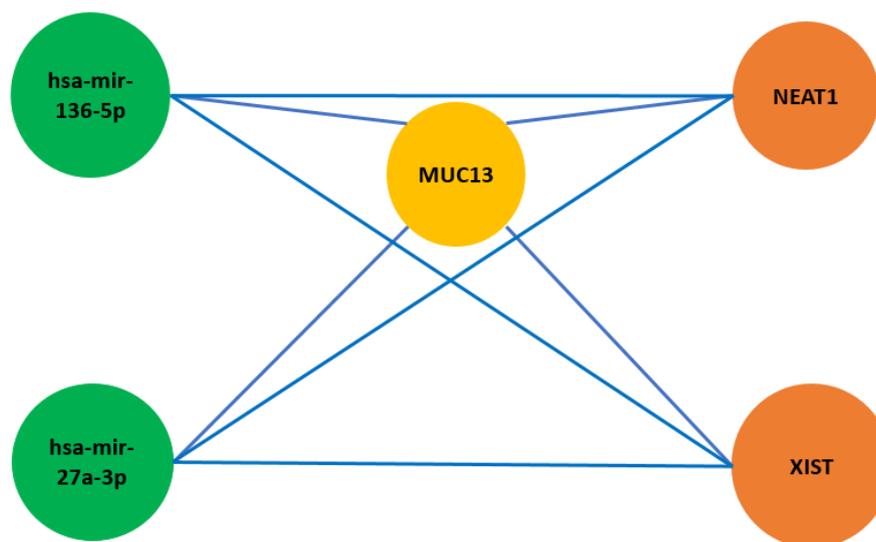
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#### Conflict of interests

The authors declare that they have no conflict of interest.

#### Author contributions

Conceived and designed the experiments: Sayyed Emad Aldin Tayyebi. Responsible for providing and quality control of the data: Sayyed Emad Aldin Tayyebi. Analysis and interpretations of the results: Sayyed Emad Aldin Tayyebi. Wrote and edited the manuscript: Sayyed Emad Aldin Tayyebi, Mahyar Heydarpour, and Hesam Dehghani.



**Figure 5.** The proposed conceptual regulatory network for the regulation of *MUC13* in colon cancer. All the predicted and validated differentially-expressed non-coding RNAs were utilized to construct these lncRNA-miRNA-mRNA axes as potential non-coding regulators of *MUC13* in colon cancer.

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